



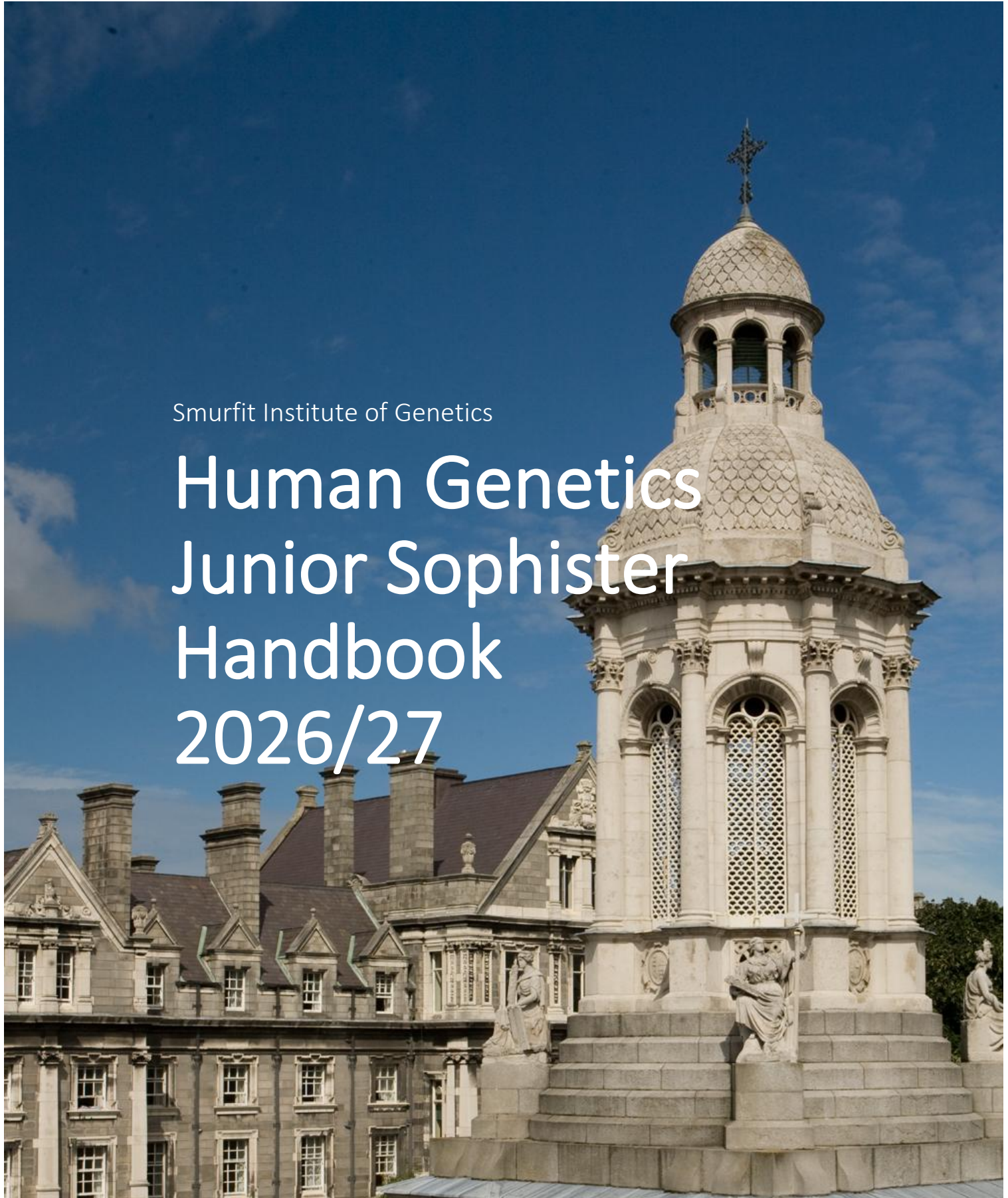
Trinity College Dublin

Coláiste na Tríonóide, Baile Átha Cliath

The University of Dublin

Smurfit Institute of Genetics

Human Genetics Junior Sophister Handbook 2026/27



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1. General Course Information

1.1 Introduction

It is a pleasure to welcome you to your Junior Sophister year in Human Genetics.

You are joining a department with a long tradition of research and teaching across a remarkably broad discipline. The study of genetics provides a framework for understanding biology at many levels - from genes, genomes and cells to development, disease, populations and evolution. Over the next two years, you will explore many of these areas through lectures, practical work, and your own research project.

Our philosophy has a strong focus on developing analytical skills. We want you to become confident in analysing evidence, thinking critically and applying your knowledge to new problems. Being able to communicate complex scientific ideas clearly is equally important, and you will have opportunities to develop both your written and oral communication skills throughout the course. I encourage you to ask questions, challenge ideas and contribute to discussions throughout your time here.

You are also now part of the Genetics Department. It is a friendly and relatively small community, and I hope you will take advantage of the opportunity to get to know the staff, researchers and other students around you. The student-run Genetics Society is an important part of departmental life, and I encourage you to get involved while also making time for the many activities available across College.

The next two years will be demanding, but they should also be rewarding and enjoyable. There will be plenty of people around you who can offer support when you need it, whether within the Department, through your personal tutor, or through the wider College support services.

I hope you enjoy your time with us, make the most of the opportunities it brings, and find plenty along the way that challenges and interests you.

With best wishes for the year ahead,
Máire Ní Leathlobhair,
JS Genetics Coordinator

1.2 Contact Details

Course Coordinator: Máire Ní Leathlobhair NLEATHLM@tcd.ie

Administrative Officer: Alicia Vega genetics@tcd.ie

Head of School: Jane Farrar Jane.Farrar@tcd.ie

DUTL: Michael John Dolan MJDOLAN@tcd.ie

School Manager: Laoise Quinn laoise.quinn@tcd.ie

Module code	Module	Coordinator	Email
MIU33302	Molecular genetics I - Regulation of gene expression	Prof Kevin Mitchell Prof Carsten Kroger	Kevin.Mitchell@tcd.ie Carsten.Kroeger@tcd.ie
GEU33301	Bioinformatics	Dr Karsten Hokamp	KAHOKAMP@tcd.ie
GEU33303	Molecular genetics II - Genome structure and dynamics	Prof Frank Wellmer	WELLMERF@tcd.ie
GEU33007	Molecular Genetics Laboratory	Dr Michael Dolan	MJDOLAN@tcd.ie
GEU33008	Analytical Genetics Laboratory	Prof Juan Pablo Labrador	LABRADOJ@tcd.ie
GEU33035	Genetic Analysis of Nervous Systems	Prof Juan Pablo Labrador	LABRADOJ@tcd.ie
GEU33045	Genomics & Systems Biology	Dr Michael Dolan	MJDOLAN@tcd.ie
GEU33055	Developmental Genetics	Prof Seamus Martin	MARTINSJ@tcd.ie
GEU33075	Evolutionary and Population Genomics	Prof Russell McLaughlin	MCLAUGR@tcd.ie
GEU33285	Science Structure, Discussion and presentation for Human Genetics	TBC	genetics@tcd.ie
GEU33215	Medical Genetics	Prof Jane Farrar	Jane.Farrar@tcd.ie
BIU33150	Biochemistry for Biosciences	Prof D. Nolan	denolan@tcd.ie
BIU33250	Introduction to Immunology & Immunometabolism	Dr Aisling Dunne	aidunne@tcd.ie
BIU33475	Basics of Neurobiology	Prof Gavin Davey	gdavey@tcd.ie

NOTE: Minor changes to that described in this JS Genetics Handbook may be undertaken during the academic year and will be notified through email to students.

1.3 Key Locations

The course will be taught in the Smurfit Institute of Genetics and Moyne Institute of Preventive Medicine.

See the maps to each institute below:

- Smurfit Institute of Genetics: <https://www.tcd.ie/Genetics/contact/>
- Moyne Institute of Preventive Medicine: <https://www.tcd.ie/Microbiology/contact/>

Teaching venues:

- LTEE3 (East End)
- Dawson (Smurfit Institute of Genetics, 2nd floor)
- Genetics meeting room (Smurfit Institute of Genetics)
- Moyne lecture theatre (Moyne Institute of Preventive Medicine)
- Moyne Lab: Microbiology Teaching Lab (Moyne Institute of Preventive Medicine)

UG study areas:

- Genetics library
- Genetics attic room

1.4 Key dates

Module	Module name	Type	Semester	Week	submission date	Submission time
BIU33150	Biochemistry for Biosciences	MCQ 1 (24%)	1	TBC	TBC	TBC
GEU33285	Human Genetics Analysis presentation	Presentations (15%)	1	TBC	TBC	TBC
GEU33007	Molecular Genetics Laboratory	Lab report 1 (50%)	1	9	Fri 30 th Oct	17:00
MIU33302	Molecular genetics I	Group presentations	1	13	Tue 24 th & Thu 26 th Nov	11:00-12:00 & 14:00-15:00
GEU33285	Human Genetics Analysis presentation	Presentations (15%)	1	13	Wed 25 th Nov	10:00-13:00
MIU33302	Molecular genetics I	Group presentations	1	13	Thurs 26 th Nov	11:00-12:00 & 15:00-16:00
MIU33302	Molecular genetics I	Group presentations	1	14	Thu 26 th Nov	15:00-16:00
GEU33075	Evolutionary and Population Genetics	MCQs (60%)	1	14	Thu 3 rd Dec	10:00-12:00
GEU33007	Molecular Genetics Laboratory	Lab report 2 (50%)	1	15	Fri 4 th Dec	17:00
BIU33150	Biochemistry for Biosciences	MCQ 2 (16%)	1	TBC	TBC	TBC
GEU33285	Science Structure, Discussion and Presentation for HGenetics	written review 4000 words (70%)	1	20	Mon 4 th Jan	9:00
GEU33301	Bioinformatics	Bioinformatics Assessment (33%)	2	26	Mon 15 th Feb	11:00-13:00
GEU33008	Analytical Genetics Laboratory	MCQ (50%)	2	26	Tuesday or Wednesday	TBC
GEU333100	Fiel trip	Presentations	2	26	25th-26th Feb	
GEU33035	Genetic Analysis of Nervous Systems	Test (50%)	2	28	Mon 8 th Mar	15:00-16:00
GEU33301	Bioinformatics	Programming Assessment (33%)	2	29	Fri 12 th Mar	10:00-12:00
BIU33250	Introduction to Immunology & Immunometabolism	MCQS (30%)	2	29	TBC	TBC
GEU33035	Genetic Analysis of Nervous Systems	class participation/essay (50%)	2	30	Mon 22 nd Mar	9:00-12:00 & 14:00-17:00
GEU33008	Analytical Genetics Laboratory	Lab report (50%)	2	29	Friday 19 th Mar	17:00
GEU33301	Bioinformatics	R programming assessment (34%)	2	32	Fri 9 th April	11:00-13:00
BIU33475	Basics of Neurobiology	continuous assessment (Literature Review) (30%)	2	33	April (TBC)	TBC

1.5 Timetable

Timetable will be circulated to students. Please be advised that the timetable might slightly vary subject to the availability of Lecturer.

It is necessary to be aware that the timetable may be subject to change if external conditions and circumstances demand this. Please be advised that the schedule might slightly vary subject to Lecturers availability. Any changes will be communicated via Blackboard announcement/email.

1.6 Behaviour in the Department

We expect high standards of personal behaviour in the Department consistent with its professional status. Please do not invite students from other Departments or friends into the Smurfit Institute, and when you are in the building, please keep the noise down. Alcohol and smoking are absolutely forbidden. Students are not permitted to go on the roofs of the buildings.

1.7 Safety

Please make sure that you have received and read the Science Faculty Safety Manual. Remember also that you are responsible for your own safety and that you have a responsibility not to endanger others by your actions. (*Refer to Appendix IX Safety Statement*)

1.8 Textbook

We recommend Introduction to Genetic Analysis by Anthony J. F. Griffiths, Susan R. Wessler, Sean B. Carroll, and John Doebley (current edition is 12th edition, 2020).

Department's web address: <http://www.tcd.ie/Genetics>

Keys – from the Genetics Executive Officer, €10 deposit.

1.9 Internships/Summer Vacation Research Experience outside Ireland

We encourage Junior Sophisters to gain experience working in a research laboratory during the summer vacation. Each year the Department awards 6 travel bursaries (4 Bill Vincent Awards, David McConnell award and James Watson award) on the basis of performance in the Senior Freshman exams, to enable students to carry out a vacation research project outside of Ireland. Prof. Matthew Campbell will advise interested students about placements in various research labs. However, it is the student's responsibility alone to arrange: air travel, Visas (where appropriate), work permits, and any insurance requirements. These arrangements should be made as far in advance as possible from the departure date – preferably in early January

1.10 Study Abroad/Erasmus

We offer our undergraduate students the opportunity to partake in Erasmus and Study Abroad programmes during their Junior Sophister year.

Students interested on a student exchange should contact the Genetics Department Office (genetics@tcd.ie) on their Senior Freshman year if their first moderatorship choice is Genetics or Human Genetics. Students will only be allowed to go on an exchange if they are expected to obtain a high 2.1 overall grade on their 2nd year (SF) passing all modules on first attempt. Students should realise that allowing a provisional exchange is dependent on finally achieving the above mentioned grade on a first attempt. A full year exchange will be considered only for students achieving a first overall grade on their first attempt.

Exchanges with other universities are organised by students and once the exchange information is provided it will be evaluated by the School of Genetics and Microbiology. A committee will evaluate the

proposal to make sure essential content is provided, the level of the modules is adequate, and all proposed modules can be attended. If approved, it will be formalised on an exchange student declaration form.

Before the exchange approval the following information will have to be provided:

- Signed Study Abroad declaration ([Appendix VI](#))
- A full list of modules available that are being considered
- Matching of international modules to Genetics/Human Genetics core modules
- Preferred modules should be highlighted for evaluation
- Information on the module content
- ECTS and ECTS equivalence for each module (some equivalence tables on [Appendix VII](#))
- Clear information of the course/year structure abroad*:
 - (i) Semester the module is taught
 - (ii) What modules are core modules and which are optional
 - (iii) Whether modules belong to different years (timetabling information will be required)

* As much information as possible, including timetabling information, if possible, will have to be provided to make sure that the proposed modules can actually be taken. Timetabling information is mandatory if modules belong to different years of a course.

We do not currently have pre-arranged agreements with universities in the Coimbra group, but some such as the University of Groningen, Netherlands, have a program that is quite compatible with Genetics and Human Genetics for an Erasmus exchange and lectures are in English. [See Coimbra group members here](#). Your first point of contact would be the person listed on the link. However, specific information about modules, timetabling etc may require you to contact specific departments within each university.

2. Scholarships and Prizes

2.1 Prizes in Human Genetics

The following prizes are awarded annually to students who have excelled during the Junior Sophister year:

- a. Leslie Bloomer Prize in Human Genetics awarded to the best qualified student in Human Genetics (based on overall JS assessment results) who in addition wishes to carry out research in the summer prior to entering the Senior Sophister year.
- b. Mcclintock Prize in Human Genetics awarded to Sophister student who has excelled in oral presentation of a subject of his/her choice within the field of Human Genetics. This prize is awarded based on presentations made on the field trip.
- c. Larry O'Hara Prize - a new prize is available and students will be provided with details on the first term

2.2 Gold Medal in Genetics

Gold medals are awarded by the Board to candidates of the first class who have shown exceptional merit in assessments for their honours bachelor degree. To be eligible, candidates must pass each year which counts towards their degree result, on the basis of a single annual attempt (which includes deferrals), and achieve the overall degree mark specified for their programme, which is set at 75 per cent or above. See <https://www.tcd.ie/academicregistry/exams/gold-medals/> for individual programme thresholds.

Various studentships, scholarships, exhibitions, and other prizes are awarded to students on the results of honour and other examinations, provided that sufficient merit is shown. Monetary awards are sent directly to prize winners unless otherwise stated under the regulations for the particular prize. For details see [Part D – Prizes and other awards](#) (Calendar 2025/26)

A book prize is awarded to each candidate obtaining an annual result of an overall first class honours grade in an honour course. These prizes are not awarded in the Senior Sophister or final year. Book prizes may be collected from the Academic Registry by the award holder in person.

3. Academic Writing

3.1 Guidance on the use of AI and Generative AI in college

The advent of commonly available artificial intelligence tools is disruptive in both positive and negative ways. Before using them in your studies, you must familiarise yourself with the College policies on their use. Unless otherwise instructed for particular modules or assessments, **the default expectation would be that you do not submit AI generated content as an attempt at an assessment.**

Below is some basic overview of the College policy on AI and GenAI. This has been taken from the more detailed policy, which is informative and wide ranging. **You are expected to have read and familiarised yourself with this policy.** https://www.tcd.ie/academicpractice/resources/generative_ai/

Artificial Intelligence (AI)

Artificial intelligence is generally understood to be a set of technologies that enable computers to perform a variety of functions usually perceived as requiring human intelligence – for example, understanding speech, recognising objects in images, composing written answers and problem reasoning. A more formal definition of an AI system from the European Union AI Act (2024) is:

...a machine-based system designed to operate with varying levels of autonomy and that may exhibit adaptiveness after deployment and that, for explicit or implicit objectives, infers, from the input it receives, how to generate outputs such as predictions, content, recommendations, or decisions that can influence physical or virtual environments[.] (EU AI Act 2024)

Generative Artificial Intelligence (GenAI)

Generative AI is the sub-area of AI, involving AI systems which generate content — for example, human dialogue, speech, images, and video. GenAI systems are capable of generating such content based on a user's request or instruction. More formally, GenAI is defined by UNESCO as **“an artificial intelligence (AI) technology that automatically generates content in response to prompts written in natural-language conversational interfaces” (UNESCO 2023).**

AI and GenAI in Trinity

As Ireland's leading university and as a world leader in AI research, Trinity recognises that AI and GenAI offer new opportunities for teaching, learning, assessment and research. We also recognise that these technologies present challenges and risks, including to academic integrity, ethics, privacy, impartiality, intellectual property and sustainability.

Acknowledging these opportunities and challenges, Trinity commits to supporting the opportunity for students and staff to become AI literate and fluent, thereby helping them to navigate and respond to the

challenges and risks of AI and GenAI in order to harness the potential of (Gen)AI to enhance teaching, learning, assessment and research – and to be prepared for future challenges as these technologies evolve. We also commit to providing ongoing resources and guidance to support students and staff to use AI and GenAI in ways that are appropriate, responsible and ethical – and to ensure that academic integrity is maintained in its usage.

College aspires to develop best practice guidelines in this area. In addition to the resources and supports that College provides and recognising that appropriate uses of AI and GenAI tools vary across academic disciplines, Schools will have some flexibility to customise their own discipline-specific practices in line with this institutional statement, other institutional policies as they develop, and national and international regulation. The College goal is to enable overall consistency in the regulation of GenAI usage, while also respecting where disciplines or degree programmes require specific restrictions in GenAI usage in assessment preparation and execution. Thus, where disciplines or degree programmes wish to refine specific regulations on student use of GenAI for learning, general as well as programme specific regulations should be communicated in the relevant discipline/degree programme handbook.

Such regulation could range from how student GenAI usage is acknowledged or cited within student assessment submissions, to prohibition of GenAI usage in the production of student assessment submissions.

[Library guidelines on acknowledging and referencing GenAI.](#)

3.2 Academic Integrity and Referencing Guide

Students of Trinity College Dublin must commit themselves to acting responsibly and ethically, embracing integrity in all actions and interactions as members of the College community, in keeping with the Council approved Statement of Integrity.

All students need to complete the [Ready Steady Write plagiarism tutorial](#), a resource developed by the Centre for Academic Practice and eLearning (CAPSL) at Trinity College Dublin, to help you understand and avoid plagiarism and develop your academic writing skills and academic integrity. It is designed so that you can view it from beginning to end or in sections and topics.

Plagiarism is using someone else's ideas, charts, concepts, or words in your assignments and using them as if they were your own, and without giving credit to the actual author. Plagiarism is considered a serious offence in Trinity and carries penalties depending on the severity of the plagiarism. To ensure that you have a clear understanding of what plagiarism is, how Trinity deals with cases of plagiarism, and how to avoid it, you will find a repository of information at <https://libguides.tcd.ie/academic-integrity>

Students may be asked for verification/validation of their work or submitted assignment via a *viva voce* or discussion/interactive session with their lecturer. Such verification would be intended to uphold quality assurance and to verify that students have met the learning outcomes. This is separate to an academic integrity meeting, which may subsequently be called if some form of Academic Integrity was suspected.

<https://www.tcd.ie/media/tcd/about/policies/pdfs/academic/assess-academic-progression.pdf>

4. Teaching and Learning

4.1 JS course structure

The Junior Sophister year is comprised of modules totaling 60 European Credit Transfer System credits (ECTS). There are 40 credits of core modules compulsory for all Genetics students, and 20 credits of open modules and Trinity Electives spread equally over two semesters in the academic year. The different possible combinations are described in the following module structure table for Genetics.

Human Genetics	
Semester 1	Semester 2
Core Modules	
MIU33302 - Molecular genetics I - Regulation of gene expression (5 credits)	GEU33301 Bioinformatics (5 credits)
GEU33007 Molecular Genetics Laboratory (5 credits)	GEU33303 - Molecular genetics II - Genome structure and dynamics (5 credits)
GEU33075 Evolutionary and Population Genetics (5 credits)	GEU33008 Analytical Genetics Laboratory (5 credits)
GEU33285 Science Structure, Discussion and Presentation for Genetics (5 credits)	GEU33035 Genetic Analysis of Nervous Systems (5 credits)
Open Modules Scenario I	
GEU33045 Genomics and Systems Biology (5 credits)	GEU33215 Medical Genetics (5 credits)
Trinity Elective (5 credits)	GEU33055 Developmental Genetics (5 credits) OR BIU33250 Introduction to Immunology and Immunometabolism (5 credits) OR BIU33475 Basic Neurobiology (5 credits)
Open Modules Scenario II	
GEU33045 Genomics and Systems Biology (5 credits)	GEU33215 Medical Genetics (5 credits)
BIU33150 Biochemistry for Biosciences (5 credits)	Trinity Elective (5 credits)
Open Modules Scenario III	
GEU33045 Genomics and Systems Biology (5 credits)	GEU33215 Medical Genetics (5 credits)
Trinity Elective (5 credits)	Trinity Elective (5 credits)

NOTE: We strongly advise you to select Open Module Scenario I and GEU33055 Developmental Genetics not to limit your Capstone Project options on your SS year.

Please consider your Open module/Trinity Elective pattern choices carefully as changes will not be permitted. If necessary, please seek advice from the relevant Course Advisor

4.2 Semester 1 Modules

MIU33302 Molecular genetics I - Regulation of gene expression

1. Module Code MIU33302

2. Module Name Molecular genetics I - Regulation of gene expression

3. Semester taught Semester 1

4. Contact Hours 29 hours

5. Module Personnel Kevin Mitchell, Carsten Kroger, Mani Ramaswami, Adrian Bracken

6. Learning Aims This module will examine the principles and processes of regulation of gene expression, in Prokaryotes and Eukaryotes. It will be anchored around the problems that organisms must solve to survive. These include modulating the cellular economy and maintaining homeostasis in response to dynamically changing internal and external states. This is crucial in microbes for adapting to environmental change and in multicellular organisms for coordinating cellular differentiation and development and regulating cellular, tissue-level and organismal physiology. The module will cover mechanisms and principles of regulation of transcription, chromatin regulation, gene regulatory motifs and networks, epigenetics, mRNA splicing, mRNA turnover, translation, non-coding RNA functions, and protein folding and localisation.

7. Module content (subject to change):

Week	Day	Time	Lecture Topic & Lecturer
3	Tue	12:00-13:00	Cellular cognition (regulating the cellular economy) (CK & KM)
3	Thu	15:00-16:00	Regulation of transcription in prokaryotes I (CK)
3	Fri	10:00-11:00	Regulation of transcription in prokaryotes II (CK)
4	Tue	12:00-13:00	Regulation of transcription in prokaryotes III (CK)
4	Thu	15:00-16:00	Group project discussion (CK & KM)
4	Fri	10:00-11:00	Gene regulatory motifs (KM)
5	Tue	12:00-13:00	Gene regulatory networks (KM)
5	Thu	15:00-16:00	Regulation of transcription in eukaryotes I (MR)
5	Fri	10:00-11:00	Regulation of transcription in eukaryotes II (MR)
6	Tue	12:00-13:00	Splicing as regulatory mechanism (MR)
6	Thu	15:00-16:00	RNA quality control (NMD) and mRNA turnover (MR)
6	Fri	10:00-11:00	Dynamical systems and landscapes (KM)
7	Tue	12:00-13:00	Tips on presentations (KM)
7	Thu	15:00-16:00	Group project meetings (CK & KM & AB & MR)
7	Fri	10:00-11:00	Group project meetings (CK & KM & AB & MR)
8	Tue	12:00-13:00	Chromatin biology I (AB)
8	Thu	15:00-16:00	Chromatin biology II (AB)
8	Fri	10:00-11:00	Chromatin biology III (AB)
9	Review week		
10	Tue	14:00-15:00	Non-coding RNAs I (CK)
10	Thu	15:00-16:00	Regulation of translation I (CK)
10	Fri	10:00-11:00	Non-coding RNAs II (KM)
11	Tue	14:00-15:00	Regulation of translation II (KM)

11	Thu	15:00-15:00	Protein folding and posttranslational modifications (KM)
11	Fri	10:00-11:00	Protein localization (KM)
12	Tue	14:00-15:00	Review / discussion (CK & KM)
13	Tue	11:00-12:00	Group Presentations (CK & KM & AB & MR)
13	Tue	14:00-15:00	Group Presentations (CK & KM & AB & MR)
13	Thu	15:00-16:00	Group Presentations (CK & KM & AB & MR)
13	Fri	10:00-11:00	Group Presentations (CK & KM & AB & MR - if needed)

NOTE: Venue Moyne lecture theatre

8. Learning Outcomes:

At the end of this module, students should:

- Have a working knowledge of the mechanisms of regulation of gene expression in Pro- and Eukaryotes.
- Understand the principles underpinning cellular cognition, homeostasis, regulation of the cellular economy, and multi-cellular development.
- Be able to deploy this knowledge and understanding to address novel problems involving these topics.
- Know how to independently research a topic in the scientific literature.
- Have experienced and reflected on the dynamics of a group project and presentation.

9. Recommended Reading List:

- Anthony J.F. Griffiths et al. Introduction to Genetic Analysis. 12th edition. New York, NY:W.H. Freeman & Company, 2020.
- Further reading on specialist topics will be provided during the presentation of the module.

10. Assessment Details:

50% continuous assessment (group project (50%) – group presentation.

50% end of semester exam (1.5-hour exam paper at the end of semester 1).

Group project:

Students will be placed into groups of four or five and given a topic related to the principles of the module. They will be expected to research the literature on this topic and prepare a short presentation to the class, using PowerPoint or similar software. The goal is for students to learn how to independently explore and assess the scientific literature, to work in a group towards a common goal, to get some experience in communicating effectively, and to deepen their understanding of the principles of the module by exploring a particular topic in detail. The group exercise will be assessed based on the presentation.

End of semester exam:

The students will be asked to answer 1 essay-type question and 8 short answer questions (3-5 sentences).

11. Module Coordinators: Kevin Mitchell (Kevin.Mitchell@tcd.ie)
Carsten Kröger (Carsten.Kroege@tcd.ie)

GEU33007 Molecular Genetics Laboratory

- 1. Module Code** GEU33007
- 2. Module Name** Molecular Genetics Laboratory
- 3. Semester taught** Semester 1
- 4. Contact Hours** 5 hours per week
- 5. Module Personnel** Mike Dolan
David Noone, Orla O'Callaghan (+ 4 demonstrators)
- 6. Learning Aims** This comprises a set of robust experiment-based projects in molecular genetics that have given us consistently good results in the Junior Sophister class environment for many years. The central theme of the module is: gene expression and its regulation.

Aims: To provide hands-on training and experience of widely used experimental strategies and techniques in molecular genetics/ molecular biology, which include: the isolation and purification of genomic and plasmid DNA; the polymerase chain reaction (PCR); using agarose and polyacrylamide gel electrophoresis to analyse DNA, RNA and proteins; genetic transformation of *E. coli*; gene cloning and analysis in plasmid vectors; *lacZ*, GUS, GFP, and immunofluorescence reporter gene assays.

7. Module content: Programme of laboratory practicals

Date	Project No.	Page	Day
	Week 3		
Wed 16 Sep	Pr 1. Exploring gene regulation in <i>E. coli</i> : the <i>lac</i> operon	p5	1 of 5
	Pr 2. Gene transfer and expression in plant cells Transformation of <i>Agrobacterium</i> with pC-TAK1	p12 p13	1 of 6
Fri 18 Sep	Pr 1. Induction of the <i>lac</i> operon: measuring <i>lacZ</i> expression Alpha-complementation and its significance in <i>lacZ</i>	p7	2 of 5
	Week 4		
Wed 23 Sep	Pr 1. PCR amplification of <i>lac</i> operon sequences	p9	3 of 5
	Pr 3. Exploring the genome and proteome of phage Growth and isolation of phage λ	p17 p18	1 of 7
	Pr 2. Purification of <i>Agrobacterium</i> transformants	p13	2 of 6
Fri 25 Sep	Pr 1. Agarose gel analysis of <i>lac</i> operon PCR products	p10	4 of 5
	Pr 3. Recovery of precipitated λ virions	p18	2 of 7
	Week 5		
Wed 30 Sep	Pr 3. Purification of λ virions on a CsCl step gradient	p18	3 of 7
	Pr 2. Liquid culture of <i>Agrobacterium</i> (pC-TAK1)	p13	3 of 6
	Pr 1. Sequence analysis of <i>lac</i> operon PCR products	p10	5 of 5
	Pr 3. Extraction of phage λ DNA	p19	3 of 7
Fri 2 Oct	Pr 2. Transformation of leaf cells via <i>Agrobacterium</i>	p13	4 of 6
	Pr 3. Restriction digestion of phage λ DNA	p19	4 of 7
	Week 6		
Wed 7 Oct	Pr 4. Creating a library of λ genes in a plasmid vector Isolation and gel analysis of pUC19 plasmid DNA	p23 p24	1 of 7

	Pr 4. Restriction digestion of pUC19	p25	1 of 7
	Pr 3. Gel analysis and processing of λ DNA digests	p20	5 of 7
Fri 9 Oct	Pr 4. Gel analysis and processing of pUC19 digests	p25	2 of 7
	Pr 4. Ligation of pUC19 and λ DNA fragments	p26	2 of 7
	Week 7		
Wed 14 Oct	Pr 3. SDS-PAGE analysis of λ virion proteins	p20	6 of 7
	Pr 4. Preparation of 'competent' <i>E. coli</i> cells and transformation with ligated pUC19- λ DNA	p27	3 of 7
Fri 16 Oct	Pr 4. Inspection of transformation plates and inoculation of miniprep cultures	p27	4 of 7
	Pr 3. Interpretation of SDS-PAGs of virion proteins	p21	7 of 7
	Week 8		
Wed 21 Oct	Pr 4. Isolation of putative pUC19- λ recombinant plasmids Restriction digestion of plasmid DNAs	p27	5 of 7
Fri 23 Oct	Pr 4. Agarose gel analysis of digested plasmid DNAs	p28	6 of 7
	Assignment of the First Lab Report		
	Week 9 – (Reading Week)		
	Week 10		
Wed 4 Nov	Pr 5. Exploring light-induced gene expression in plants 1. Differential regulation of rRNA gene expression	p30-31	1 of 4
	Pr 2. Detection of transgenes in regenerated leafy shoots a. Detection of the <i>Kan^r</i> gene by PCR b. Detection of the GUS gene via reporter activity	p14	5 of 6
Fri 6 Nov	Pr 5. Agarose gel analysis of rRNAs	p31	2 of 4
	Pr 2. Agarose gel analysis of <i>Kan^r</i> PCR products	p15	6 of 6
	Pr 4. Sequence analysis of pUC19- λ clones	p28	7 of 7
	Week 11		
Wed 11 Nov	Pr 5. Exploring light-induced gene expression in plants: 2. Regulation of RUBISCO synthesis	p32-33	3 of 4
	Pr 5. Exploring light-regulated gene expression in plants: 3. Regulatory properties of the <i>RbcS</i> promoter	p34-36	3 of 4
Fri 13 Nov	Pr 5. Interpretation of RUBISCO gels Discussion: Regulation of gene expression by light	p33	4 of 4
	Assignment of the Second Lab Report		
	Week 12		
Wed 18 Nov	Pr 6. Exploring reporter gene expression in the mouse brain with immunofluorescence (TBC)		
Fri 20 Nov			
	Lab Report writing (16 – 20 Nov)		

NOTE: Labs will run on:

- Wed 14:00-17:00 - Venue: Biolab3
- Fri 14:00-16:00 - Venue Moyne Lab: Microbiology Teaching Lab

8. Learning Outcomes: On successful completion of the module, students will be able to:

- isolate and purify genomic and plasmid DNA;
- assemble the reagents required to amplify DNA using the polymerase chain reaction (PCR);
- analyse DNA and RNA using agarose gel electrophoresis;
- make protein extracts and analyse them using SDS-polyacrylamide gel electrophoresis;
- transform *E. coli* and *Agrobacterium* using plasmid DNA;
- grow, isolate and purify bacteriophage;
- clone DNA fragments in plasmid vectors;
- use *Agrobacterium* to introduce genes into the plant nucleus;
- perform *lacZ*, GUS, GFP and immunofluorescence reporter gene assays.

9. Recommended Reading List: published primary research papers and reviews

10. Assessment Details:

Students prepare two Laboratory Reports of equal weighting.

Students are notified of (1) the first Laboratory Report during Week 6 on October 9th, and (2) the second report during Week 11 on November 13th.

The first report is due at 17:00 Irish Time, October 30th 2026.

The second report is due at 17:00 Irish Time, December 4th 2026.

11. Module Coordinator: Mike Dolan MJDOLAN@tcd.ie

GEU33075 Evolutionary and Population Genetics

1. Module Code GEU33075

2. Module Name Evolutionary and Population Genetics

3. Semester taught Semester 1

4. Contact Hours Lectures 24 hours; tutorials: 4 hours

5. Module Personnel Lara Cassidy, Russell McLaughlin, Ross McManus, Marco Capodiferro

6. Learning Aims This module provides an in-depth exploration of genetic variation, from its origins to its evolutionary consequences. The information in DNA is not always transmitted accurately from one generation to the next. DNA sequences can change spontaneously by the process of mutation and inaccurate DNA repair, resulting in genetic variation (polymorphism) within populations. Variable sites at different positions in the genome get shuffled into new combinations by the process of genetic recombination that occurs during sexual reproduction. Whether a particular allele survives for a long time in a population or goes extinct depends on the evolutionary forces acting on the population. If a new allele is advantageous to the population, Darwinian natural selection will tend to increase its frequency in the population; alternatively, if the new allele is disadvantageous natural selection will tend to eliminate it. However, selection is only one of several evolutionary processes that change allele frequencies within populations over generations. In this module, students will learn about the origin of genetic variation, its distribution within populations and long-term changes brought about by evolutionary processes.

7. Module content

Week	Day & Time	Lecture Topic & Lecturer
3	Mon 14 th Sept 10:00-11:00	Introduction to evolutionary and population genetics (McLaughlin)
3	Tue 15 th Sep 14.00-15.00	RMM L1: Spectrum and mechanisms of DNA mutation I (McManus)
3	Tue 15 th Sep 15.00-16.00	RMM L2: Spectrum and mechanisms of DNA mutation II (McManus)
3	Fri 18 th Sep 11:00-12:00	RMM L3: Exogenous and structural causes of mutation (McManus)
3	Fri 18 th Sep 12:00-13:00	RMM L4: Mutation and health (McManus)
4	Mon 21 st Sep 09:00-10.00	RMcl L1: Genetic variation and its detection I (McLaughlin)
4	Mon 21 st Sep 10.00-11.00	RMcl L2: Genetic variation and its detection II (McLaughlin)
4	Mon 21 st Sep 14.00-15.00	RMcl L3: Alleles and genotypes: Hardy-Weinberg equilibrium (McLaughlin)
4	Mon 21 st Sep 15.00-16.00	RMcl L4: Alleles and genotypes: inbreeding (McLaughlin)
4	Fri 25 th Sept 11:00-12:00	RMcl L5: Effective population size (McLaughlin)
4	Fri 25 th Sept 12:00-13:00	RMcl L6: Genetic drift, fixation F-statistics and population structure (McLaughlin)
5	Mon 28 th Sep 09:00-10.00	RMcl L7: Linkage disequilibrium (McLaughlin)
5	Mon 28 th Sep 10:00-11.00	RMcl L8: Applied population genetics: complex traits and GWAS (McLaughlin)
5	Mon 28 th Sep 14.00-15.00	RMcl L9: Applied population genetics: population structure and PCA (McLaughlin)
5	Mon 28 th Sep	Extra slot

	15.00-16.00	
6	Mon 5 th Oct 09:00-10.00	LC L1: Evolutionary change in sequences: measuring evolution (Cassidy) Blackboard – prerecorded
6	Mon 5 th Oct 10.00-11.00	LC L2: Evolutionary change in sequences: patterns and models (Cassidy) Blackboard – prerecorded
6	Mon 5 th Oct 14.00-15.00	LC L3: Evolutionary change in sequences: the molecular clock (Cassidy) Blackboard – prerecorded
6	Mon 5 th Oct 15.00-16.00	LC L4: Evolutionary change in sequences: molecular phylogenetics (Cassidy) Blackboard – prerecorded
6	Tue 6 th Oct 14:00-15:00	LC L5: Evolutionary change in sequences: applied phylogenetics (Cassidy) Blackboard – prerecorded
6	Tue 6 th Oct 15:00-16:00	Extra slot
7	Mon 12 th Oct 09:00-10.00	LC L6: Genome evolution: gene duplication (Cassidy) Blackboard - prerecorded
7	Mon 12 th Oct 10.00-11.00	LC L7: Genome evolution: concerted evolution (Cassidy) Blackboard - prerecorded
7	Mon 12 th Oct 14.00-15.00	LC L8: Genome evolution: whole genome duplication (Cassidy) Blackboard - prerecorded
7	Mon 12 th Oct 15.00-16.00	LC L9: Genome evolution: gene loss and base composition evolution (Cassidy) Blackboard - prerecorded
7	Tue 13 th Oct 14:00-15:00	LC L10: Genome evolution: Transposition (Cassidy) Blackboard - prerecorded
7	Tue 13 th Oct 15:00-16:00	Tutorial – Lectures 1-5 (Capodiferro)
8	Mon 19 th Oct 09:00-10.00	Extra slot
8	Mon 19 th Oct 10.00-11.00	Extra slot
8	Mon 19 th Oct 14.00-15.00	Extra slot
8	Mon 19 th Oct 15.00-16.00	Extra slot
8	Tue 20 th Oct 14:00-15:00	Extra slot
8	Tue 20 th Oct 15:00-16:00	Tutorial – Lectures 6-10 (Capodiferro)
9	Review week	
14	Thu 3 rd Dec 10:00-12:00	MCQ Exam venue (MAC LAB)

NOTE: Venue LTEE3 Groups for tutorials will be facilitated by course coordinator.

Description of each Lecture

- **Introduction to evolutionary and population genetics** (Cassidy)
This lecture introduces molecular evolution and provides historical context, including ideas presented by Lamarck, Weissmann and Darwin. Core principles, including selection and neutral mutations, are also introduced.
- **Spectrum and mechanisms of DNA mutation I** (McManus)

- **Spectrum and mechanisms of DNA mutation II** (McManus)
These two lectures explore the different ways that DNA can change (mutate) between individuals, leading to genetic variation within a population and a substrate on which evolution acts. Types of DNA mutation are described, along with the molecular biological mechanisms bringing about these changes.
- **Exogenous and structural causes of mutation** (McManus)
This lecture extends our view of the origin of mutation beyond molecular mechanisms to include exogenous (eg environmental) factors and structural changes.
- **Mutation and health** (McManus)
The consequences of mutation on human health are discussed.
- **Genetic variation and its detection I** (McLaughlin)
- **Genetic variation and its detection II** (McLaughlin)
These two lectures describe in detail some core methodologies and technologies used in the identification and study of genetic variation in DNA. Methods include polymerase chain reaction, electrophoresis, Sanger sequencing, whole-genome SNP genotyping and next-generation sequencing. Technologies are also compared and contrasted for their relative advantages and constraints in the context of conducting population genetic studies on genetic variation.
- **Alleles and genotypes: Hardy-Weinberg equilibrium** (McLaughlin)
In this lecture, we define the expected relationship between allele and genotype frequencies in an idealized population, and the interpretation when observation deviates from expectation. Statistical methods for assessing the probability of observed data under a null model are detailed, and the conclusions that can be drawn from the results of these tests are discussed.
- **Alleles and genotypes: inbreeding** (McLaughlin)
The concept of inbreeding (mating between related individuals) is introduced and our model for expected genotype frequencies under various inbreeding scenarios is updated. This forms the basis of ensuing lectures.
- **Effective population size** (McLaughlin)
In this lecture we state how populations are modelled as idealized populations with effective sizes that can – sometimes counterintuitively – be far from actual census sizes. We discuss the various parameters that affect effective population size and their consequences in the population, including resilience in agriculture and animal breeding.
- **Genetic drift, fixation, F-statistics and population structure** (McLaughlin)
Here we explore genetic drift (the random long-term change in allele frequencies) and the ultimate long-term fate for all genetic variation: either loss or fixation. We explore the role of parameters such as population size on defining the rate of fixation of an allele in a population. We then set a statistical framework for the analysis of variance of genotype frequencies in a population and their use in defining the fixation index, a useful tool for identifying genetic structure (non-random patterns) in populations.
- **Linkage disequilibrium** (McLaughlin)
This lecture introduces the new concept of linkage disequilibrium – the statistical correlation of adjacent alleles – and its uses in modern genomics.
- **Applied population genetics: complex traits and GWAS** (McLaughlin)
- **Applied population genetics: population structure and PCA** (McLaughlin)
In these final three lectures, we apply some of the techniques and methods that have been discussed in previous lectures to understand the genetic basis of traits in a population and genomic descriptors of population differentiation. We explore the design and typical execution

of a genome-wide association study (GWAS), from its motivation to the statistical considerations in controlling false positives and technical bias. We qualitatively discuss the application of principal component analysis (PCA) in delineating population structure and controlling GWAS, and finish with a tour of modern, population-scale genome sequencing studies such as the UK Biobank.

- **Evolutionary change in sequences: measuring evolution** (Cassidy)
In this lecture we learn how biological entities are compared on the molecular level and introduce the concept of molecular homology. In part two, we discuss the evolutionary forces that cause related sequences to diverge through time. We define a substitution event and introduce the molecular clock and neutral theory.
- **Evolutionary change in sequences: patterns and models** (Cassidy)
In this lecture we consider evolution as a series of substitution events and introduce substitution models, which can be used to describe the patterns and rates of sequence evolution. We discuss the assumptions of these models and violations to them. The concept of functional constraint is introduced, as well as the Ka/Ks ratio method for detecting the action of purifying and positive selection on protein-coding sequences.
- **Evolutionary change in sequences: the molecular clock** (Cassidy)
The concept of the molecular clock and its applications are explored in more detail. We learn how to calibrate a molecular clock with an outgroup and use it to estimate species divergence times. The core assumption of the clock - a constant rate of substitution - is explored and the causes of non-constant rates of evolution across lineages are introduced.
- **Evolutionary change in sequences: molecular phylogenetics** (Cassidy)
This lectures explains the construction of phylogenetic trees from sequence data to describe the evolutionary relationships between species. The aims of the field of phylogenetics are outlined and students are given a refresher in phylogenetic terminology and the concept of reproductive isolation. Methods for reconstructing and rooting trees are described in turn. Methods for assessing tree reliability are also given.
- **Evolutionary change in sequences: applied phylogenetics** (Cassidy)
This lecture provides examples of real-world applications of phylogenetics. This includes the characterization of the last common ancestor of all life and the closest living relatives of cetaceans (whales and dolphins). Viral phylogenetics is also introduced.
- **Genome Evolution: Gene Duplication** (Cassidy)
This lecture answers the question: 'where do new genes come from?' The mechanisms and evolutionary consequences of gene duplication are discussed. In part two, several examples of gene duplication driving phenotypic evolution are provided (globins, olfactory receptors and primate opsins).
- **Genome Evolution: Concerted evolution** (Cassidy)
This lecture defines and explores concerted evolution, the process which results in the non-independent evolution of genes that are closely related in sequence. Duplicated genes within a species may remain similar due to the horizontal spread of mutations by unequal crossing over and gene conversion. Both mechanisms are discussed in turn. The concerted evolution of rRNA repeats, globin genes in primates, and ribonucleases in ruminants are given as examples.
- **Genome Evolution: whole genome duplication** (Cassidy)
The concept of polyploidy is defined and its incidence in the natural world explored. The mechanisms and evolutionary consequences of true polyploidy are discussed. In part two, we learn about the process of diploidization, whereby a polyploid species reverts to a diploid state through structural rearrangements of the genome and gene loss.

- **Genome Evolution: gene loss and base composition evolution** (Cassidy)

This lecture covers two distinct topics. In part one, we learn about the mechanisms and consequences of gene loss. We ask why a single copy gene may be lost from a species and cover several examples of gene loss in humans and other species. In part two, we learn about base composition - the proportions of the four bases within DNA. We ask why GC content differs across species and discuss both mutational and selection-based hypotheses for this variation. We also look at GC and AT skews on the two strands of the DNA helix, as well as codon usage bias.

- **Genome Evolution: transposition (mobile genetic elements)** (Cassidy)

Transposition is discussed in this lecture – the process by which genetic elements are transferred “horizontally” between organisms rather than the more conventional “vertical” inheritance of genetic material from parent to offspring. Types of transposable elements are introduced and their prevalence in nature outlined. The evolutionary consequences of transposition are also discussed.

8. Learning Outcomes

On completion of this module, students will be able to:

- Describe and explain the origin of genetic variation through mutation
- Describe and explain the consequences of genetic variation in health and disease
- Identify suitable technological approaches to the detection and study of different types of genetic variation
- Describe the relationship between population genotype and allele frequencies under panmixia and inbreeding
- Define effective population size (N_e), identify population parameters that influence N_e and describe practical consequences of low N_e even in large census populations
- Specify the long-term evolutionary trajectories of alleles under genetic drift and define fixation
- Define and quantitatively describe population structure using F-statistics
- Define linkage disequilibrium and describe its use in population genomics
- Explain the design of genome-wide association studies for complex traits and their use of population genetic principles to control bias
- Discuss modern population-scale genome sequencing efforts to understand human health and disease
- State the neutral theory of molecular evolution
- Describe the nature and consequences of evolutionary change in DNA and protein sequences, including codon usage bias
- Define the relationship between mutation accumulation and evolutionary time and its use in the molecular clock
- Construct phylogenetic trees based on evolutionary relationships between related sequences
- Define homolog, paralog, ortholog and ohnolog and describe concerted evolution of paralogous genes within species
- Describe large-scale evolutionary changes to genetic sequences, including transposition, gene and genome duplication and gene loss

9. Recommended Reading List

Introduction to Genetic Analysis, 11th ed. (Griffiths, Wessler, Carroll, Doebley) – chapters 15-18
Principles of Population Genetics, 4th ed. (Hartl & Clark)

10. Assessment Details

This module will be graded through an invigilated inhouse MCQ examination (60%) on 3rd Dec (MAC Lab) and One 1.5-hour exam paper at the end of semester 1 (40%).

For the MCQ, each student will receive a random set of questions that capture the entire breadth of the module. These questions will be designed to test for both quick recall and reasoning skills. Students may be asked to solve mathematical problems, interpret tabular data and phylogenetic diagrams. There will be no negative marking.

11. Module Coordinator: Russell McLaughlin (MCLAUGR@tcd.ie)

GEU33085 Science Structure, Discussion and Presentation for Genetics

1. Module Code GEU33285

2. Module Name Science Structure, Discussion and Presentation for Human Genetics

3. Semester taught 1

4. Contact Hours 12

5. Module Personnel Adrian Bracken, Dan Bradley, Matthew Campbell, Mike Dolan, Jane Farrar, Seamus Martin, Kevin Mitchell, Juan Pablo Labrador, Russell Mc Laughlin, Mani Ramaswami, Frank Wellmer, Máire Ní Leathlobhair, Basma Baccouche, Avril Reddy, Gergo Porkolab, Magdalena Karpinska, Anahit Hovhannisyan, Marco Capodiferro, Conor Rossi.

6. Learning Objectives: The module has the following learning objectives: (1) discussion of the design and implementation of genetic analysis of biological phenomena relevant to human genetics; (2) discussion of genetic analysis in cell and animal models - their strengths and weaknesses; (3) discussion of mathematical genetics; (4) discussion of how to research, design and write a literature review on a human genetics subject; (5) to write a 4,000 literature review on an assigned human genetics topic; (6) discussion of science communication: how to assemble and present a talk on a human genetics topic. Students will make a 10 minute presentation on a paper/review each week (week 4-7) within the topic of the tutorial.

(7) Students will make a 15 minute presentation on their literature review at the annual field-trip

7. Module content: Programme of tutorials

Week		Programme of Tutorials	GEU33285
3	Wed 16 th Sept 10:00-11:00	Module introduction (Mitchell)	
3	Wed 16 th Sept 11:00-12:00	How to make an oral presentation (Mitchell)	
3	Thu 17 th Sept 12:00-13:00	Review I: How to write a literature review on a genetics/human genetic topic (Mc Laughlin) VENUE LTEE3	
3/4	One to one meeting with supervisor	Review II: Supervisor and student meet to discuss the topic and how to structure the review. (student to reach out supervisor and organise meeting)	
4	Wed 23 rd Sept 9:00-12:00	Human Genetic Analysis I: In search of human disease genes (Baccouche)	
5	Wed 30 th Sept 9:00-12:00	Human Genetic Analysis II: Understanding gene function (Reddy)	
6	One to one meeting with supervisor	Review III: Supervisor and student meet to discuss review structure produced by the student.	
6	Wed 7 th Oct 9:00-12:00	Human Genetic Analysis III: Studying mechanisms (Porkolab)	
7	Wed 14 th Oct 9:00-12:00	Human Genetic Analysis IV: Emerging methodologies in elucidating the molecular basis of human disease (Karpinska)	
9		Reading Week	
10	Wed 4 th Nov 10:00-12:00	Mathematical Genetics I: Networks/graph theory (Rossi, Hovhannisyan & Capodiferro)	
11	Wed 11 th Nov 10:00-12:00	Mathematical Genetics II: Game theory: Evolution of co-operation/altruism (Rossi, Hovhannisyan & Capodiferro)	
12	Wed 18 th Nov 12:00-13:00	Mathematical Genetics III: Bayesian statistics (Rossi, Hovhannisyan & Capodiferro)	
13	Wed 25 th Nov 10:00-13:00	Mathematical Genetics Presentations	
19	Mon 4 th Jan 9am	Assignment written literature review submission	

NOTE: Venue: Dawson

- Week 3: **How to make an oral presentation:** Discussion of how to prepare a talk, focusing on the topic, judging your audience, slide preparation and the general do's and don'ts of preparing and giving an oral presentation. Discussion will be centred on "*Giving a seminar: Suggestions for Graduate Students*" by John Roth.
- Week 3 **Review I: How to write a literature review on a genetics/human genetics topic:** Discussion under the following headings: Topic / focus, structure of review, themes, progression of ideas, literature survey, search databases, references.
- Week 3/4 **Review II: Discussion of the review topic assigned to each student and review with the supervisors.** This is arranged by the supervisor / student at their mutual convenience.
- Week 4 **Human Genetic Analysis I: In search of human disease genes.** TBC
- Weeks 5 **Human Genetic Analysis II: Understanding gene function** TBC
- Weeks 6 **Human Genetic Analysis III: Studying mechanisms** TBC
- Week 7 **Human Genetic Analysis IV: Emerging methodologies in elucidating the molecular basis of human disease** TBC
- Week 10 **Mathematical Genetics I** - Networks/graph theory
- Week 11 **Mathematical Genetics II** - Game theory: Evolution of co-operation/altruism.
- Week 12 **Mathematical Genetics III** - Bayesian statistics.
- Week 13 **Mathematical Genetics IV** - Mathematical Genetics presentations.

8. Learning Outcomes: Students will have the following skills: (1) to research and write a scientific literature review; (2) to research, prepare and give a scientific oral presentation; (3) to design and perform a genetic analysis of a biological phenomenon relevant to human genetics; (4) to choose an appropriate model organism in which to study specific biological phenomena; (5) to perform mathematical genetic analysis.

9. Recommended Reading List:

Tutorials in weeks 4-7 will be based on the following reviews:

Nat Rev Genet. 2015 16(5): 275–284 Genetic linkage analysis in the age of whole-genome sequencing.

Jurg Ott, Jing Wang and Suzanne M. Leal

Adv Exp Med Biol 2020;1236:1-38. A History of Mouse Genetics: From Fancy Mice to Mutations in Every Gene. María J García-García

How to make a scientific presentation: "*Giving a seminar: Suggestions for Graduate Students*" by John Roth.

Reading material for the Mathematical genetics tutorials will be presented during the first tutorial.

10. Assessment Details:

- 15% Human Genetic Analysis Presentations
- 15% Mathematical Genetics Presentations
- 70% Written review, 4,000 words (deadline, Monday of week 19, January 4th 9am)

11. Module Co-ordinator TBC

GEU33045 Genomics & Systems Biology

1. Module Code GEU33045 ([Open Module](#))

2. Module Name Genomics & Systems Biology

3. Semester taught 1

4. Contact Hours 24

5. Module Personnel Mike Dolan, Adrian Bracken, Matthew Dorman, Kenneth Mok

6. Learning Aims The aim of this module is to equip students with a comprehensive understanding of the methods used in the fields of genomics, proteomics and metabolomics and how these methods are used for basic research, biotechnology, agriculture and medicine. To this end, several applications from work in diverse organisms (bacteria, fungi, plants, animals including humans) in addition to specific diseases and disorders (Schizophrenia and Cancer) will be presented. The module further introduces students to the field of systems biology and outlines how systems biology differs from the classic reductionist approach used in biology.

7. Module content:

Week	Day & Time	Lecture Topic & Lecturer
3	Mon 14 th Sept 17:00-18:00	Introduction to Genomics and Systems Biology (Dolan)
3	Tue 15 th Sept 17:00-18:00	History of DNA Sequencing I (Dolan)
4	Mon 21 st Sept 17:00-18:00	History of DNA Sequencing II: The Human Genome Project (Dolan)
4	Tue 22 nd Sept 17:00-18:00	Modern Day DNA sequencing I: 2nd Generation Sequencing Technologies (Dolan)
5	Mon 28 th Sept 17:00-18:00	Modern Day DNA sequencing II: 3rd Generation Sequencing Technologies (Dolan)
5	Tue 29 th Sept 17:00-18:00	Reference Sequences and Alignment (Dolan)
6	Mon 5 th Oct 17:00-18:00	Population Genomics (Dolan)
6	Tue 6 th Oct 17:00-18:00	Transcriptomics: Revealing Gene Expression (Dolan)
6	Wed 7 th Oct 17:00-18:00	How to Find Genes (Dolan)
7	Mon 12 th Oct 17:00-18:00	Functional Genomics: Regulatory Genome and non-coding DNA (Dolan)
7	Tue 13 th Oct 17:00-18:00	Single-cell and Spatial Transcriptomics I (Dolan)
8	Mon 19 th Oct 17:00-18:00	Single-cell and Spatial Transcriptomics II (Dolan)
8	Tue 20 th Oct 17:00-18:00	<i>Revision of material, discussion and answering student questions (Dolan) – tutorial</i>
9		Review week
10	Mon 2 nd Nov 17:00-18:00	Bacterial genomes and comparative genomics (Dorman)
10	Tue 3 rd Nov 17:00-18:00	Introduction to the 'epigenome': histone and DNA modifications (Bracken)
10	Wed 4 th Nov 17:00-18:00	Proteomics: Identify/characterise/quantify; Mass Spec and other technologies (Mok)
11	Mon 9 th Nov 17:00-18:00	Functional genomics in bacteria (Dorman)
11	Wed 11 th Nov 17:00-18:00	Quantitative proteomics; clinical proteomics (Mok)

12	Mon 16 th Nov 17:00-18:00	Methods to analyse the epigenome; the ENCODE project 1 (Bracken)
12	Tue 17 th Nov 17:00-18:00	Extra slot
12	Wed 18 th Nov 17:00-18:00	Interaction/affinity proteomics; metabolomics introduction (Mok)
13	Mon 23 rd Nov 17:00-18:00	Methods to analyse the epigenome; the ENCODE project 2 (Bracken)
13	Wed 25 th Nov 17:00-18:00	Metabolomics technologies (Mok)
14	Mon 30 th Nov 17:00-18:00	Cancer profiling and using genomic information for the development of cancer therapies (Bracken)
14	Tue 1 st Dec 17:00-18:00	Revision of material, discussion and answering student questions (all lecturing staff)
15		Revision Week
16		Assessment Week

NOTE: Venue LTEE3

8. Learning Outcomes: Upon successful completion of this module, students will be able to describe experimental approaches used in the fields of genomics, proteomics and metabolomics. They will understand how to leverage these methods to analyze complex biological systems and questions. Students will be able to evaluate the applications of these techniques in biological sciences and discuss case studies involving specific diseases and disorders. Finally, they will be able to differentiate between systems biology and traditional reductionist approaches in biology.

9. Recommended Reading List: none

10. Assessment Details: One 1.5-hour exam paper at the end of semester 1

11. Module Coordinator Mike Dolan MJDOLAN@tcd.ie

Last year schedule and module descriptor. New timetable and details will be provided on Blackboard. Please use this information as a guide

1. Module Code BIU33150 (Open Module)

2. Module Name Biochemistry for Biosciences

3. Semester taught Semester 1

4. Contact Hours 20 hours lectures online (each ~ 50 mins in total), plus 2 in person tutorial sessions

5. Module Personnel Amir Khan (AK), K.H. Mok (KHM), Vincent Kelly (VK), Martin Caffrey (MC), Andrei Budanov (AB), Derek Nolan (DN), Emma Creagh (EC), Aisling Dunne (AD), Daniela Zisterer (DZ).

6. Learning Aims This module follows on from the biochemistry/cell biology component of the "Molecules to Cells" BIU22201 module of year 2. The aim is to provide Junior Sophister students of other disciplines with the grounding in biochemistry necessary to (i) understand biology at a molecular level, (ii) form a mechanistic view of biological processes and (iii) appreciate the pathobiochemical basis of disease. The module covers four major themes in biochemistry: Proteins and Nucleic Acids, Membranes, Cytoskeleton and Signalling. The module will be assessed through a combination of in course assessment and an individual end of term exam.

7. Module content: Programme of lectures

Lecture	Lecture Topic & Lecturer
Part A - Proteins and Nucleic Acids	
1	Amino acids and peptide bond (AK)
2	Structures, motifs and folds (AK)
3	Structure and mechanism: serine proteases (AK)
4	Spectrophotometry of biomolecules (KHM)
5	Protein folding and pathologies (KHM)
6	The proteome (KHM)
7	DNA, chromatin & the nucleus (VK)
8	RNA structure, folds & function (VK)
Part B – Membranes	
9	An introduction to cellular and model membranes (MC)
10	Membrane composition and therapeutic approaches (MC)
11	The Synthetic & Assembly Mechanisms for Membrane Proteins that Form Specific Topologies (DN)
12	Membrane transport of small molecules, specificity, mechanisms, energy Coupling (DN)
Part C – Cytoskeleton	
13	Structure of tubulin and microtubules and the Assembly / Disassembly of Microtubular Structures (EC)
14	Microtubular motors, types, mechanism of movement, regulation, physiological roles (EC)
15	Introduction to actin and the actin cytoskeleton (DN)
16	F-actin nucleation & pathologies associated with actin cytoskeleton (DN)
Part D – Signalling	
17	Introduction to cell signalling & GPCRs (EC)
18	G-Protein coupled Receptor (GPCR) regulation (EC)
19	Receptor tyrosine kinases (RTKs)-PDGF and EGF (DZ/AD)
20	RTK signalling – PKB and PDK1 (DZ/AD)

Lectures schedule and time table

All lectures will be delivered online as follows with the accompanying lecture notes on blackboard : Module BIU33150. Lectures will be released 2 per week starting on week 1 of the teaching term (Monday 9th Sept). Subsequent lectures will be released 2 per week each following Monday. Lectures are usually subdivided into part A , C etc on BB. These can vary in length but combined will give 50 min overall for the lecture topic. So these are not multiple lectures, they are smaller parts of the same lectures.

Sample MCQs and exam Qs are available on line in BB.

Date of lects (on BB)	Lect No	Staff	Upload to BB by
1 Mon 11/09	1-2	AK	11/09
2 Mon 18/9	3-4	AK/KHM	18/09
3 Mon 25/9	5-6	KHM	25/09
4 Mon 2/10	7-8	VK	2/10
5 Mon 9/10	9-10	MC	9/10
6 Mon 16/10	11-12	DN	16/10
7 Reading week			
8 Mon 30/10	13-14	EC	30/10
9 Mon 6/11	15-16	DN	6/11
10 Mon 13/11	17-18	EC	13/11
11 Mon 20/11	19-20	AD/DZ	20/11

(1-11 week of teaching term)

8. Learning Outcomes: On completion of this module, the student will be able to:

- Recall and comprehend key knowledge and concepts of the hierarchy of polypeptide structure and the forces that stabilize the three-dimensional shape of proteins
- Explain the link between a protein structure and its biological activity, and with appropriate examples, how human diseases arise from a deviation in structure
- Appreciate the principles of spectrophotometry and its applications to biomolecules
- Understand the concept of the proteome and its importance in disease
- Integrate key concepts about nucleic acid structure and function
- Demonstrate an understanding of the biochemical processes of nucleic acids in the cell
- Recall and integrate key knowledge and concepts concerning the role of lipids in membrane structure and function
- Describe how model membranes are formed and their applications
- Describe how an understanding of membrane composition and structure can be used in the design of vaccines, antibiotics and beta-blockers.
- Demonstrate a knowledge of the biosynthesis of membrane proteins, including the mechanisms of insertion and transport to their various locations.
- Explain the types of membrane transport and how this process is coupled to energy and assayed.
- Describe the structure of microtubules, their assembly and disassembly and their polarity.
- Describe the structure of microtubule motors and the processes of directed vesicle transport and cytoplasmic streaming.
- Describe the structure of monomeric actin and how it is assembled into filaments
- Explain how actin nucleation is linked to pathological states.

- Describe the general principles of G-protein coupled receptor (GPCR) signalling and its regulation, the initial discovery of G-proteins linked to cyclase, the functional effects of cAMP and the activation of GPCR-linked signal-activated phospholipases.
- Discuss Receptor Tyrosine Kinase (RTK) signalling and details of MAP kinase cascades, using PDGF and EGF as examples. Explain RTK and PI3K pathways in the context of PKB (Akt) and PDK1 signalling.
- Describe the principles of steroid hormone receptor signalling mechanisms.

9. Recommended Reading List: A reading list will be given out by lecturers during the module.

10. Assessment Details: 60% End of year examination, 40% in course assessed.

In course assessment: Two online MCQ assessments.

The MCQs will be structured as follows:

There will be two MCQs per lecture, giving 40 MCQs for the course.

The first MCQ will be after reading week and will cover the first 12 lectures (so 24 MCQs) representing 24% of the marks for the module.

The second MCQ will be in week 12, i.e. the last week of the term and will cover the material contained in the eight lectures after reading week. There will be 16 MCQs representing 16% of the marks for the course.

11. Module Coordinator: Dr D Nolan

denolan@tcd.ie

4.3 Semester 2 Modules

GEU33100 Field Trip

1. Module Code GEU33100

2. Module Name Field Trip

3. Semester taught 2

4. Contact Hours 14

5. Module Personnel Adrian Bracken, Dan Bradley, Matthew Campbell, Lara Cassidy, Máire Ní Leathlobhair, Mike Dolan, Jane Farrar, Seamus Martin, Kevin Mitchell, Juan Pablo Labrador, Aoife Mc Lysaght, Russell McLaughlin, Mani Ramaswami, Frank Wellmer.

6. Learning Objectives: The field trip takes place over two days on the week 27 (25th – 26th Feb 2027). Hilary Term). It is a great opportunity for staff and students to meet scientifically and socially in an informal setting. Each student is expected to present a short (10- minute) presentation on the topic of their allocated Literature Review.

7. Module content:

Week		Lecture Topic & Lecturer
24	Mon 8 th Feb 10:00-11:00	Field trip tutorial (Wellmer) LTEE3
26	Thurs 25 th Feb 10:00-18:30	Student presentations
26	Fri 26 th Feb 10:00-15:00	Student presentations

8. Learning Outcomes: At the end of this module, students should be able to deliver a professional presentation on a scientific subject.

9. Recommended Reading List: Individually suggested by supervisors and student personal research.

10. Assessment Details: 10 min presentation on week 26 (25th – 26th Feb 2027). Attendance and presentation are **compulsory**.

11. Module Coordinators: Frank Wellmer WELLMERF@tcd.ie

GEU33303 Molecular genetics II - Genome structure and dynamics

1. Module Code GEU33303
2. Module Name Molecular Genetics II
3. Semester taught 2
4. Contact Hours 23 (5 ECTS)
5. Module Personnel Prof. Frank Wellmer, Prof Matthew Dorman
6. Learning Aims The aim of this module is to introduce students to fundamental concepts in Molecular Genetics. By focusing on plant and microbial genetics, we want to highlight the overlap between these seemingly disparate biological systems. The microbial genetics component will focus on critical regulatory aspects of the gene expression machinery (transcription and translation), and genome replication (DNA replication, homologous recombination, mutagenesis and DNA repair). In relation to plant genetics, students will be introduced to major topics such as the structure and evolution of the nuclear genome, the importance of model plants, including *Arabidopsis thaliana*, light-regulated gene expression, hormone receptors and signal transduction systems. The evolutionary origins of plant cell organelles (chloroplasts and mitochondria) via endosymbiosis involving ancestral microbes will also be explored as will be the many facets of plant-microbe interactions including plant immunity and symbiosis.
7. Module content:

Week	Date & Time	Lecture Topic & Lecturer
21	Tue 19 th Jan 11:00-12:00	DNA replication I: Linear and circular genomes; replication strategies; origins of replication (Dorman)
21	Tue 19 th Jan 14:00-15:00	DNA replication II: regulation of initiation at oriC; role of DnaA and hemimethylation (Dorman)
21	Thurs 21 st Jan 15:00-16:00	Homologous recombination I: Holliday, Meselson-Radding and Szostak models (Dorman)
22	Tue 26 th Jan 14:00-15:00	Homologous recombination II: RecA, SSB, RecBCD, RuvA, B, C; Horizontal gene transfer (Dorman)
22	Thurs 28 th Jan 15:00-16:00	DNA Mutagenesis: chemical, deamination, base-analogues, chromosomal integration through homologous DNA, transposons (Dorman)
23	Tue 2 nd Feb 11:00-12:00	DNA Repair systems: Mismatch base repair; Base excision repair; nucleotide excision repair; SOS response; Non-homologous end-joining; Error Prone DNA Polymerases (Dorman)
23	Tue 2 nd Feb 14:00-15:00	<i>Tutorial: revision of material, discussion and answering student questions (Dorman)</i>
23	Thurs 4 th Feb 15:00-16:00	Introduction: why plant research is important (Wellmer)
24	Tue 9 th Feb 11:00-12:00	Models for plant research and nuclear genome structure (Wellmer)
24	Thurs 11 th Feb 15:00-16:00	Evolution of the nuclear genome and sex determination in plants (Wellmer)
25	Thurs 18 th Feb 15:00-16:00	Discovery and mutagenic potential of transposable elements (Wellmer)
27		Review Week
28	Wed 10 th Mar 16:00-17:00	Regulation of transposon activity (Wellmer)

28	Thurs 11 th Mar 14:00-15:00	The plastid genome and plastid evolution (Wellmer)
29	Wed 17 th Mar 16:00-17:00	Bank Holiday
29	Thurs 18 th Mar 14:00-15:00	The genetic toolbox for analyzing plant biology: the making of transgenic plants (Wellmer)
29	Thurs 18 th Mar 15:00-16:00	<i>Tutorial: revision of material, discussion and answering student questions (Wellmer)</i>
30	Wed 24 th Mar 16:00-17:00	The genetic toolbox for analyzing plant biology: genetic screens (Wellmer)
30	Thurs 25 th Mar 14:00-15:00	The genetic toolbox for analyzing plant biology: mutant analyses (Wellmer)
30	Thurs 25 th Mar 15:00-16:00	Understanding signal transduction in plants (Wellmer)
31	Wed 31 st Mar 16:00-17:00	Understanding the response pathway for light (Wellmer)
31	Thurs 1 st Apr 14:00-15:00	Understanding the response pathway for the hormone ethylene (Wellmer)
31	Thurs 1 st Apr 15:00-16:00	Plant-microbe interactions (Wellmer)
32	Thurs 8 th Apr 14:00-15:00	Plant-pathogen interactions: an evolutionary arms race (Wellmer)
32	Thurs 8 th Apr 15:00-16:00	<i>Tutorial: revision of material, discussion and answering student questions (Wellmer)</i>

8. Learning Outcomes: Upon successful completion of this module, students will be able to describe the critical regulatory features of the gene expression systems in prokaryotic microbes and eukaryotic cell organelles (chloroplasts and mitochondria). They will understand how DNA is replicated, how it can be recombined via homologous recombination, altered by mutagenic processes, and repaired. Students will also have acquired knowledge of key topics in plant genetics, such as light-regulated gene expression, hormone receptors and signal transduction, and the genetic basis of plant immunity.

9. Recommended Reading List: Anthony J.F. Griffiths et al. Introduction To Genetic Analysis. 12th edition. New York, NY :W.H. Freeman & Company, 2020.

Further reading on specialist topics will be provided during the presentation of the module.

10. Assessment Details: One 1.5-hour exam paper at the end of semester 2

11. Module Coordinators: Frank Wellmer (wellmerf@tcd.ie)
Matthew Dorman (dormanmj@tcd.ie)

GEU33301 Bioinformatics

- 1. Module Code** GEU33301
- 2. Module Name** Bioinformatics
- 3. Semester taught** Semester 2
- 4. Contact Hours** 39 Hours
- 5. Module Personnel** Dr Karsten Hokamp (KH), Dr Carsten Kröger (CK), Dr Máire Ní Leathlobhair (MNL), Dr Fiona Roche (FR)
- 6. Learning Aims** This module is taught in combination with the Microbiology department and covers web-based bioinformatics, Python programming and a data handling component. Lectures will be held in computer labs to enable a hands-on approach. The bioinformatics component provides a practical introduction to the use of commonly used bioinformatic databases and tools with a focus on web-based applications. Students will become familiar with accessing biological sequence databases and exploring various sequence analysis tools to understand evolutionary relationships and how this can help to draw protein functional and structural inferences. The Python programming component introduces students to computer programming in Python using bioinformatics-related examples and exercises. It will be carried out within an internal JupyterLab environment. The data handling part contains a biolab component in which samples will be prepared for whole-genome sequencing (WGS). The combined lectures and practicals cover basic techniques for data processing and working with the statistical software R.

This module runs for 14 x 2 hours (combined lecture and practical sessions) plus a 5-hour lab practical and 3 x 2-hour in-class assessments in Semester 2.

Students are required to bring their own laptop for the practical sessions.

As learning aims of this module students will

- learn how to search a range of biological databases
- understand how proteins are annotated and classified
- get to know which tools can be used to explore unknown sequences of interest
- master the concept of sequence alignment and homology searching
- understand the process of programming
- acquire basic Python programming skills
- gain a thorough understanding of next-generation sequencing data
- be familiar with the steps and tools required to process, map and visualise NGS data
- become familiar with the specifics of NGS data analysis
- understand how to enhance NGS results with annotation data
- isolate genomic DNA from *E. coli* bacteria
- prepare a DNA library for Nanopore sequencing
- carry out WGS analysis of a bacterial genome

7. Module content: Programme of lectures and practicals

Week	Day & Time	Lecture Topic & Lecturer
21	Mon 18 th Jan 14:00-16:00	Bioinformatics 1 – Biological Databases (FR, 2 hours)
21	Fri 22 nd Jan 11:00-13:00	Bioinformatics 2 – Protein Resources & Function Prediction (FR, 2 hours)
22	Mon 25 th Jan 14:00-16:00	Bioinformatics 3 – Pairwise and Multiple Sequence Alignments (FR, 2 hours)
22	Fri 29 th Jan 14:00-16:00	Bioinformatics 4 – Homology Searching using BLAST (FR, 2 hours)
23	Fri 5 th Feb 14:00-16:00	Programming 1 – Operators, Variables and Loops (KH, 2 hours)
24	Mon 8 th Feb 14:00-16:00	Programming 2 – Input/Output, Branching (KH, 2 hours)
24	Fri 12 th Feb 14:00-16:00	Programming 3 – Lists, Tuples, Sets (KH, 2 hours)
25	Mon 15 th Feb 11:00-13:00	Bioinformatics Assessment (FR, 2 hours)
25	Mon 15 th Feb 14:00-16:00	Programming 4 – Dictionaries (KH, 2 hours)
25	Fri 19 th Feb 11:00-13:00	Programming 5 – Functions, System Commands (KH, 2 hours)
Review week		
28	Tue 9 th Mar 11:00-12:00 & 14:00-18:00	Biolab Practical (CK, 5 hours) ** Venue Biolab 1
28	Thu 11 th Mar 9:00-11:00	Data Handling 1 – Using R for bioinformatics: Part I (MNL, 2 hours)
28	Fri 12 th Mar 11:00-13:00	Programming Assessment (KH, 2 hours)
29	Thu 18 th Mar 11:00-13:00	Data Handling 2 – Using R for bioinformatics: Part II (MNL, 2 hours)
29	Fri 19 th Mar 11:00-13:00	Data Handling 3 – Graphs & Data Visualization in R (MNL, 2 hours)
30	Thu 25 th Mar 11:00-13:00	Data Handling 4 – Working with Notebooks in R (MNL, 2 hours)
31	Thu 1 st Apr 11:00-13:00	Data Handling 5 – Introduction to Machine Learning in R (MNL, 2 hours)
32	Fri 9 th Apr 11:00-13:00	R Assessment (MNL, 2 hours)

NOTE: Venue LTEE3 and MAC Lab.

Biolab 1 Carsten Kroger session on Tue 9th March

Description of each Lecture:

Bioinformatics 1 - Biological Databases (FR, 2 hours) This lecture covers how bioinformatics data are stored and organised with a focus on resources provided at NCBI and UCSC. Students will learn about the different types of data and tools found within these resources, specifically the retrieval of gene/transcript information and approaches to interactively explore biological data in the context of the genome.

Bioinformatics 2 - Protein Resources and Function Prediction (FR, 2 hours) This lecture describes how protein sequence data are stored, annotated and classified. Students will learn about computational methods used for predicting protein function and be introduced to protein resources, such as UniProt,

AlphaFold and InterPro.

Bioinformatics 3 - Pairwise and Multiple Sequence Alignment (FR, 2 hour) This lecture introduces the concept of sequence alignment, the process of comparing two sequences to determine if they are evolutionarily related to one another. Students will gain an understanding of both pairwise and multiple sequence alignment and learn of its many applications including its role in inferring function and structure and the identification of genetic variants from sequence comparison.

Bioinformatics 4 - Homology Searching using BLAST (FR, 2 hours) This lecture will explore the concept of homology searching using the BLAST algorithm, which compares single sequences against a database of sequences to search for significantly similar sequences. PSI-BLAST will also be introduced for the identification of more distant homologs.

Programming 1 – Operators, Variables and Loops (KH, 2 hours) This lecture covers mathematical operators, string variables, string formatting, as well as built-in functions and methods for strings. This is followed by the use of ‘while’ and ‘for’ loops for repeated application of programming steps.

Programming 2 – Input/Output, Branching (KH, 2 hours) This lecture deals with ways of reading experimental data from files into a Python script and how to store results generated by a script in a file. It also introduces ways of making decisions in a Python script through branching. With this additional skill set students will be able to write more elaborate scripts and tackle processing of sequence data in Fasta format.

Programming 3 – Lists, Tuples, Sets (KH, 2 hours) Lists are a very common feature in data science. They are represented in Python through various types of iterable variables, which are covered in this lecture together with their built-in functions and methods.

Programming 4 – Dictionaries (KH, 2 hours) This lecture is dedicated to dictionaries, a collection of key-value pairs, which enables students to implement a DNA translation script.

Programming 5 – Functions, System Commands (KH, 2 hours) With the introduction of functions, the repertoire of programming skills is expanded so that scripts can be written more efficiently. System commands will enable the execution of external programs from within a script.

Practical – Isolation of gDNA and Nanopore Sequencing (CK, 5 hours) In this practical, the students will learn how to isolate genomic DNA from *Escherichia coli* bacteria and how to quality-control the DNA integrity to be suitable for whole genome sequencing. One DNA library will be prepared for Oxford Nanopore sequencing.

Data Handling 1 – Using R for Bioinformatics: Part I (MNL, 2 hours) This lecture will introduce students to how the statistical software R can be used to manipulate and analyse data. Students will learn about data structures, data manipulation and subsetting, as well as functions and packages in R.

Data Handling 2 – Using R for Bioinformatics: Part II (MNL, 2 hours) Building on Part I, this lecture will explore more complex data manipulation methods and learn about specialized bioinformatics packages in R. The session will also cover statistical analysis, enabling students to apply these skills to their own research projects.

Data Handling 3 - Graphs & Data Visualization in R (MNL, 2 hours) This lecture will cover basic plotting in base R, data visualization with ggplot2 and introductory data visualization theory.

Data Handling 4 - Working with Notebooks in R (MNL, 2 hours) This lecture covers the use of R Markdown Notebooks as an integrated way to carry out and share data analysis. Students will learn about the structure of R Notebooks, how to use Markdown to format text, and how to create and customize code blocks within Notebooks.

Data Handling 5 – Introduction to Machine Learning in R (MNL, 2 hours) This lecture will introduce students to the basics of machine learning using R. Students will learn about key machine learning concepts and techniques, including supervised and unsupervised learning.

8. Learning Outcomes: On successful completion of the module students should be able to:

- MLO1. Query a range of bioinformatic databases
- MLO2. Apply tools to investigate unknown sequences
- MLO3. Carry out sequence alignment and homology searching
- MLO4. Visualise biological data through a genome browser
- MLO5. Approach programming tasks in a structured way
- MLO6. Write Python scripts following good coding practice
- MLO7. Integrate external programs and use functions within Python scripts
- MLO8. Solve entry-level bioinformatics problems using Python scripts
- MLO9. Assess the quality of NGS data
- MLO10. Apply bioinformatics tools for processing NGS data
- MLO11. Visualise NGS data through genome browser resources
- MLO12. Carry out core NGS downstream analyses resulting in genome assemblies
- MLO13. Integrate external annotation data with analysis results through Python scripts
- MLO14. Competency to isolate bacterial DNA and to prepare a Nanopore sequencing reaction

9. Recommended Reading List:

The Biostar Handbook, 2nd Edition (<https://www.biostarhandbook.com/>)

Bioinformatics, 4th Edition, John Wiley and Sons Ltd, by Andreas D. Baxevanis, Gary Bader, David Wishart

Bioinformatics and Functional Genomics, 3rd Edition, 2015, Wiley Blackwell, Jonathan Pevsner

A Critical Guide to BLAST, TK Attwood

<https://f1000research.com/documents/7-1435>

How to Think Like a Computer Scientist, Learning with Python 3 (RLE), 2012 edition, by Peter Wentworth, Jeffrey Elkner, Allen B. Downey, and Chris Meyers

(<https://openbookproject.net/thinkcs/python/english3e/>)

- 10. Assessment Details:**
- (1) A bioinformatics exam 33%
 - (2) a Python programming exam 33%
 - (3) an R programming exam 34%.

In case of an overall fail mark for this module, the failed components need to be repeated during the reassessment period.

11. Module Coordinator: Dr Karsten Hokamp kahokamp@tcd.ie

GEU33035 Genetic Analysis of Nervous Systems

1. Module Code GEU33035

2. Module Name Genetic Analysis of Nervous Systems

3. Semester taught Semester 2

4. Contact Hours 30

5. Module Personnel Juan Pablo Labrador, Mani Ramaswami

6. Learning Aims The module is focused on understanding how experimental genetics are used to manipulate genes in organisms to address problems in biology. Areas covered are 1) Experimental Genetics: structure and conservation of genes, nature of mutations and their effects on protein structure and function, model organisms in genetic research and experimental manipulation of animal genomes. 2) Developmental Neurogenetics: the purpose and design of genetic screens, genetic analysis of neurogenesis and genetic analysis of axon guidance 3) Behavioral Genetics: cell organization and methods of cell biology, cell biology of neurons and synapses, creation and use of molecular reporters of specific gene or cell activity, methods to study nervous systems, sensory circuits, sensation; transduction; perception; coding; behavior, learning and memory, sleep and circadian rhythms.

7. Module content: Program of lectures

Week	Day & Time	Lecture Topic & Lecturer
21	Tue 19 th Jan 12:00-13:00	Experimental Genetics - Introduction - Structure and conservation of genes, nature of mutations and their effects on protein structure and function (Labrador)
21	Wed 20 th Jan 15:00-16:00	Experimental Genetics - Dominance and germ line transmission (Labrador).
21	Thu 21 st Jan 9:00-10:00	Experimental Genetics - Transgenesis and the Gal 4 system (Labrador). LTEE1
21	Thu 21 st Jan 11:00-12:00	Experimental Genetics - Experimental manipulation of animal genomes. (Labrador)
21	Thu 21 st Jan 12:00-13:00	Developmental Genetics - The purpose and design of genetic screens (Labrador)
22	Tue 26 th Jan 12:00-13:00	Developmental Genetics – Mapping Genes (Labrador)
22	Wed 27 th Jan 15:00-16:00	Developmental Genetics - From phenotypes to genes, midline screen (Labrador)
22	Thu 28 th Jan 11:00-12:00	Developmental Genetics - From genes to pathways, modifiers screens (Labrador)
22	Thu 28 th Jan 12:00-13:00	Neurogenetics/ Behavioral Genetics Overview (Ramaswami)
23	Tue 2 nd Feb 12:00-13:00	Action Potentials; Neural Excitability; Channelopathies, (Ramaswami)
23	Wed 3 rd Feb 15:00-16:00	Channelopathies, Synaptic Transmission (Excitation, Inhibition, Modulation (Ramaswami)
23	Thu 4 th Feb 11:00-12:00	Vision, taste and smell (genetics/biology) and principles of neural coding (Ramaswami)
23	Thu 4 th Feb 12:00-13:00	Temperature sensing and coding (Ramaswami)
24	Tue 9 th Feb 12:00-13:00	Cellular Basis for Memory (Ramaswami)

24	Wed 10 th Feb 15:00-16:00	Genetic analysis of memory in Drosophila (Ramaswami)
24	Thu 11 th Feb 11:00-12:00	Circadian Rhythms and Overview of Behavioral Genetics(Ramaswami)
24	Thu 11 th Feb 12:00-13:00	TBC
25	Wed 17 th Feb 15:00-16:00	TBC
25	Thu 18 th Feb 11:00-12:00	TBC
25	Thu 18 th Feb 12:00-13:00	TBC
27		Review week
28	Mon 8 th Mar 15:00-16:00	Final written test
29	Mon 15 th Mar 16:00-17:00	TBC
30	Mon 22 nd Mar 9:00-12:00	Presentations
30	Mon 22 nd Mar 14:00-17:00	Presentations

NOTE: Venue LTEE3 // **week 21 Thursday 9:00-10:00 LTEE1**

Lecture times are subject to change within the hours reserved for the module on the above timetable.

8. Learning Outcomes: Upon successful completion of this module, students will be able to understand and describe how model organisms are used in genetic research and common technologies and methods employed to genetically modify organisms. Students should also understand the basis of genetic screens and mapping. They will be able to explain epistasis through the analysis of different genetic interactions in neurogenesis and axon guidance. Students will become familiar with the cell biology of neurons and synapse as well as methods to probe synaptic activity. Students will also learn about circuitry underlying perception.

9. Recommended Reading List: Anthony J.F. Griffiths; Susan R. Wessler; Sean B. Carroll; John Doebley. Introduction To Genetic Analysis. New York, NY :W.H. Freeman & Company, 2015

10. Assessment Details: Continuous assessment:
Test (50%) - week 28
Class participation/Poster presentation (50%) - Week 30

11. Module Coordinator Juan Pablo Labrador labradoj@tcd.ie

GEU33008 Analytical Genetics Laboratory

- 1. Module Code** GEU33008
- 2. Module Name** Analytical Genetics Laboratory
- 3. Semester taught** Semester 2
- 4. Contact Hours** 5 hours per week
- 5. Module Personnel** Juan Pablo Labrador
- 6. Learning Aims**

This module is a practical module that introduces the fundamentals of genetic analysis and the use of *Drosophila melanogaster* as a genetic model organism. The module will cover different aspects of model organisms handling and segregation. Virtual crosses are employed to understand Mendelian genetics and non-Mendelian inheritance including segregation, recombination, gene mapping, lethal genes and sex-linked inheritance.

- 7. Module content:** Programme of lectures

Week	Day & Time	Lecture Topic & Lecturer
21	Mon 18 th Jan 16:00-18:00	Drosophila husbandry, identification of phenotypes, setting up crosses
	Tue 19 th Jan 15:00-17:00	Mendelian inheritance tutorial. P-elements tutorial
	Wed 20 th Jan 16:00- 18:00	Monohybrid, dihybrid crosses (Labrador)
22	Tue 26 th Jan 15:00-17:00	Drosophila husbandry, identification of phenotypes, setting up crosses
	Wed 27 th Jan 16:00- 18:00	Non-Mendelian inheritance tutorial, Lethal mutations. (Labrador) Short Quiz 1: Monohybrid/dihybrid crosses
23	Tue 2 nd Feb 15:00-17:00	Segregation in Drosophila, identification of males carrying P-elements
	Wed 3 rd Feb 16:00- 18:00	Lethal mutations , Sex linked inheritance. (Labrador) Short Quiz 2: Lethal mutations
24	Tue 9 th Feb 15:00-17:00	Segregation in Drosophila - fly husbandry
	Wed 10 th Feb 16:00- 18:00	Sex linked inheritance , Linkage, recombination and mapping. Linkage and mapping tutorial (Labrador) Short Quiz 3: Sex linked inheritance
25	Tue 16 th Feb 15:00-17:00	Segregation in Drosophila analysis of crosses
	Wed 17 th Feb 16:00- 18:00	Linkage and mapping (Labrador) Short Quiz 4: Linkage, Recombination and mapping
26	Mon 22 nd Feb 14:00-17:00	Fly husbandry -Set up crosses if required
	Tue 23 rd Feb 15:00-17:00	Review Mendelian and non-Mendelian inheritance (Labrador)
	Wed 24 th Feb 16:00- 18:00	Final Quiz: Mendelian and non-Mendelian inheritance, lethal mutations, linkage and mapping (date TBC)
27		Reading Week
29	Mon 15 th Mar 14:00-17:00	Segregation in Drosophila – Review of results P-element mapping in Drosophila Tutorial (Labrador) Fly husbandry

30	Tue 23 rd Mar 15:00-16:00	Analysis and review of P-element results Fly husbandry (Labrador)
31	Wed 31 st Mar 16:00- 18:00	TBC

NOTE: Venue MAC LAB and BIOLAB3 - Experiment weeks may be subject to change as we are working with live animals. An updated experiment, MCQs and lab report timetable will be provided at the start of the module.

8. Learning Outcomes: Upon successful completion of this module, students will be able to understand how model organisms are used in genetic research in a laboratory setting. Students should be able to set-up crosses and plan experiments using *Drosophila melanogaster*. Students will be able to design crosses and analyse ratios in the progeny. Mendelian inheritance ratios through the analysis of monohybrid, dihybrid and trihybrid crosses involving two or 3 chromosomes should be understood. Students will appreciate non-Mendelian inheritance involving sex linked loci or linked loci in autosomes or sex chromosomes.

9. Recommended Reading List:

Anthony J.F. Griffiths; Susan R. Wessler; Sean B. Carroll; John Doebley. Introduction To Genetic Analysis. New York, NY :W.H. Freeman & Company, 2015

Greenspan, R.J. (2004). Fly pushing: the theory and practice of *Drosophila* genetics, 2nd ed. (Cold Spring Harbor, N.Y.: Cold Spring Harbor Laboratory Press).

10. Assessment Details: MCQs/Final MCQ 50% Week 26.

Lab report 50% submission deadline Friday week 29 (TBC) 5pm

11. Module Coordinator Juan Pablo Labrador labradoj@tcd.ie

GEU33055 Developmental Genetics

1. Module Code GEU33055 (Open Module)

2. Module Name Developmental Genetics

3. Semester taught Semester 2

4. Contact Hours 24

5. Module Personnel Prof. Seamus Martin, Prof. Adrian Bracken, Prof. Frank Wellmer

6. Learning Aims This module aims at introducing students to fundamental concepts in developmental genetics and to experimental approaches that are used to study development. To this end, the module takes a comparative approach: the development of different organisms (insects, vertebrates, plants) will be examined to demonstrate differences and commonalities in the genetic mechanisms controlling morphogenesis. Students will be introduced to important developmental control mechanisms, including morphogens, spatial signaling via asymmetric distribution of signaling molecules, signal transduction cascades that culminate in differential gene expression and the activation of homeotic selector genes which dictate the specification of biological structures such as limbs and wings. We will also explore the factors governing cell division, a key biological process that is used to generate huge numbers of new cells during development as well as for tissue turnover and repair, and we will examine how cell division is precisely regulated via the 'cell division cycle'. The module will also introduce students to stem cell biology and will examine how stem cells are programmed to undergo growth and differentiation.

7. Module content:

Week	Day & Time	Lecture Topic & Lecturer
21	Wed 20 th Jan 11:00-12:00	Intro to development and developmental genetics-Overview (Martin)
21	Wed 20 th Jan 14:00-15:00	Pluripotent Stem Cells (Bracken)
21	Thu 21 st Jan 14:00-15:00	Multipotent Stem Cells 1 (Bracken)
22	Wed 27 th Jan 11:00-12:00	The cell cycle 1 (Martin)
22	Wed 27 th Jan 14:00-15:00	Multipotent Stem Cells 2 (Bracken)
22	Thu 28 th Jan 14:00-15:00	Cellular Reprogramming (Bracken)
23	Wed 3 rd Feb 11:00-12:00	The cell cycle 2 (Martin)
23	Wed 3 rd Feb 14:00-15:00	The cell cycle 3 (Martin)
23	Thu 4 th Feb 14:00-15:00	The cell cycle 4 (Martin)
24	Wed 10 th Feb 11:00-12:00	Introduction to <i>Drosophila</i> development and embryogenesis (Martin)
24	Wed 10 th Feb 14:00-15:00	<i>Drosophila</i> development: anterior-posterior axis specification (Martin)
24	Thu 11 th Feb 14:00-15:00	<i>Drosophila</i> development: role of maternal effect genes (Martin)

25	Wed 17 th Feb 11:00-12:00	<i>Drosophila</i> development: activation and function of gap genes (Martin)
25	Wed 17 th Feb 14:00-15:00	<i>Drosophila</i> development: activation and function of pair-rule genes (Martin)
25	Thu 18 th Feb 14:00-15:00	<i>Drosophila</i> development: activation and function of segment polarity genes (Martin)
26	Wed 24 th Feb 11:00-12:00	<i>Drosophila</i> development: dorso-ventral axis specification (Martin)
26	Wed 24 th Feb 12:00-13:00	<i>Drosophila</i> development: homeotic selector genes and segment identity (Martin)
26	Wed 24 th Feb 14:00-15:00	Tutorial: revision of material, discussion and student questions (Martin, Bracken)
27		Review week
28	Wed 10 th Mar 11:00-12:00	Organogenesis: genetic control of vertebrate limb development (Wellmer)
28	Wed 10 th Mar 14:00-15:00	Organogenesis: genetic control of vertebrate limb development II (Wellmer)
30	Wed 24 th Mar 11:00-12:00	Organogenesis: Genetic control of <i>Drosophila</i> eye development (Wellmer)
30	Wed 24 th Mar 14:00-15:00	Principles of plant development (Wellmer)
31	Wed 31 st Mar 11:00-12:00	Stem cells, cell fate determination and organogenesis in plants (Wellmer)
31	Wed 31 st Apr 14:00-15:00	Tutorial: revision of material, discussion and student questions (Wellmer)

NOTE: Venue LTEE3

8. Learning Outcomes: Upon successful completion of this module, students will be able to describe fundamental mechanisms underlying the control of development in animals and plants. They will understand how cell division is tightly regulated in higher organisms via a dedicated set of enzymes (called the cyclins and cyclin-dependent kinases) and how deregulated cell division can lead to cancer. They will have acquired knowledge to outline key events during development including embryogenesis and the formation of organs and will be able to describe how these events are regulated at a molecular level. Students will be familiar with the biology of stem cells and how these cells are programmed to undergo growth and differentiation. They will be able to describe experimental approaches that are used by researchers to dissect developmental processes and mechanisms.

9. Recommended Reading List:

Anthony J.F. Griffiths; Susan R. Wessler; Sean B. Carroll; John Doebley. Introduction To Genetic Analysis. New York, NY :W.H. Freeman & Company, 2015.

10. Assessment Details: One 1.5-hour exam paper at the end of semester 2

11. Module Coordinator Prof. Seamus Martin Email: MARTINSJ@tcd.ie

GEU33215 Medical Genetics

1. Module Code GEU33215 (Open Module)

2. Module Name Medical Genetics

3. Semester taught Semester 2

4. Contact Hours 18

5. Module Personnel Jane Farrar, Russell McLaughlin,

6. Learning Objectives The module provides an introduction to core concepts in medical genetics highlighting the importance and power of genetic information in the era of genomic medicine and the impact of such information for all of us. Learning objectives include: (1) discussion of the genetic basis of single gene disorders (Mendelian) and complex disorders; (2) overview of the history of medical genetics; (3) insights into key developments in medical genetics up to 2025 including state-of-art technologies and novel innovative therapies; (4) discussion of the key technologies and methodologies currently used to elucidate the genetic basis of human traits; (5) discussion of the individualisation of medicine and the important roles of genetic information in disease diagnosis, prognosis and the design and choice of therapy. In summary, the module provides an introduction to:

- The genetic basis of mendelian & complex disorders
- Genetic technologies & methodologies used to elucidate the genetic basis of human traits
- The exploitation of genomic data in diagnosis, prognosis & treatment of disease
- The individualization of medicine using genetic information

7. Module content: Programme of lectures

Week	Day & Time	Lecture Topic & Lecturer
28	Mon 8 th Mar 17:00-18:00	Importance of medical genetics and history of developments in the field. (Farrar)
28	Wed 10 th Mar 17:00-18:00	Introduction to complex traits. (McLaughlin)
28	Fri 12 th 16:00-17:00	Quantitative traits and complex disorders. (McLaughlin)
28	Fri 12 th 17:00-18:00	Variance components and heritability (McLaughlin)
29	Mon 15 th Mar 17:00-18:00	Overview of spectrum of Mendelian and multifactorial disorders. (Farrar)
29	Tue 16 th Mar 17:00-18:00	Identification of disease genes and disease mutations. (Farrar)
29	Fri 19 th Mar 16:00-17:00	Dominance, additivity and interaction. (McLaughlin)
29	Fri 19 th Mar 17:00-18:00	Selection on complex traits and the liability scale for complex disorders. (McLaughlin)
30	Mon 22 nd Mar 17:00-18:00	Variant interpretation and ACMG guidelines. (Farrar)
30	Tue 23 rd Mar 17:00-18:00	New trends in medical genetics. (Farrar)
30	Wed 24 th Mar 17:00-18:00	Selection on complex traits and the liability scale for complex disorders. (McLaughlin)
31	Tue 30 th Mar 17:00-18:00	Introduction to pharmacogenomics. (Farrar)
31	Wed 31 st Mar	Mapping complex disease genes I: GWAS and polygenic risk. (McLaughlin)

	17:00-18:00	
31	Fri 2 nd Apr 16:00-17:00	Genetic variation in cytochrome P450s and influence on drug response. (Farrar)
31	Fri 2 nd Apr 17:00-18:00	Tracking on polygenic pharmacogenetic traits - GWAS, high throughput analysis. Actionable pharmacogenetic traits in clinical practice (Farrar)
32	Mon 5 th Apr 17:00-18:00	Tutorial (Farrar)
32	Tue 6 th Apr 17:00-18:00	Mapping complex disease genes II: missing heritability and rare variants (McLaughlin)
32	Wed 7 th Apr 17:00-18:00	Tutorial (McLaughlin)
32	Fri 9 th Apr 16:00-17:00	Extra slot
32	Fri 9 th Apr 17:00-18:00	Extra slot

NOTE: Venue LTEE3.

Description of Lectures:

Jane Farrar

History and importance of medical genetics: Topics include disease prevalence (congenital, Mendelian and multifactorial disorders including cancer), the practical value of genetic information *inter alia* the role of genetic counseling and potential of emerging new therapeutics based on knowledge of disease etiology.

Overview of spectrum of Mendelian and multifactorial disorders: Topics include phenotypes associated with autosomal dominant, recessive and X-linked disorders, emphasising the clinical significance of many of these inherited disorders with many being distressing, serious and life-threatening. The genetics underpinning multifactorial disorders and methodologies utilised to elucidate these genetic factors is also discussed.

Identification of disease genes and disease mutations. An overview of the development and utilisation of methodologies to identify disease causing genes and mutations. Application of genome sequencing in the era of genomic medicine.

New trends in medical genetics: In the era of whole genome sequencing (WGS) interpretation of novel genetic variants is key; innovative methods for interpretation are discussed. Use of the American College of Medical Genetics (ACMG) guidelines in clinical genetics. Targeted therapies directed by genetic information – current status for gene therapy, gene editing etc.

Introduction to pharmacogenomics. Genetic variation between individuals greatly influences drug response, drug efficacy and / or drug toxicity and indeed is relevant to all of us - on average every human carries a number of actionable pharmacogenetic variants. Such genetic information is leading to the individualisation of medicine and has enormous ramifications for medicine and drug development. An introduction to methods used to identify genetic variants for monogenic and polygenic pharmacogenetic traits will be discussed. Genes encoding drug metabolising enzymes (DMEs) will be introduced including Phase I and Phase II DMEs. For example, the significant effects of genetic variants in the thiopurine methyltransferase (TPMT) gene (a Phase II DME) on response to major chemotherapeutic and immunosuppressant drugs will be outlined.

Genetic variation in cytochrome P450s and drug response: The family of genes encoding cytochrome P450 enzymes (Phase I DMEs) will be introduced including CYP2C9 and CYP2D6, among others. Genetic variants in these DME genes influence metabolism of drugs, including commonly used medications

such as warfarin and codeine, among others. A summary of the studies underpinning the now actionable pharmacogenomic outcomes associated with warfarin and codeine therapies will be provided. Genetic information can be utilised to target the optimal therapy at the right dose to the correct patient.

Tracking on polygenic pharmacogenetic traits - GWAS, high throughput analysis. Actionable pharmacogenetic traits in clinical practice. Methods used to identify genetic factors that influence risk of an adverse drug response between individuals will be reviewed. Parallels between pharmacogenomic studies and use of GWAS, WGS to decipher genetic determinants of complex disorders will be highlighted. While pharmacogenetic traits underpin much of the variance between individuals in response to medications, genetic variants in pharmacogenes can also influence disease risk. Many DMEs metabolise not only exogenous compounds (therapeutics) but also endogenous compounds, and therefore it is not surprising that genetic variants in these genes can contribute to disease risk - mounting evidence that genetic variants in pharmacogenes can impact disease risk will be outlined. Genetic variants that can cause catastrophic drug responses will be discussed, as will how these are becoming actionable in clinical practice. An overview will be provided of pharmacogenetic traits that have obtained FDA/EMA backing with actionable outcomes in clinical practice. The barriers that have hindered translation of pharmacogenetic information from the laboratory into clinical practice will be discussed.

Russell McLaughlin

Introduction to complex traits: In this lecture we define the concept of complex traits, framed against classical Mendelian genetics and alternative models of genetic architecture. This lecture also explores the problem of defining phenotypes in the presence of clinical and genetic heterogeneity.

Quantitative traits and complex disorders: This lecture sets the statistical framework for following lectures, defining basic population parameters including mean, variance and distribution, using a worked example of human height. The liability scale is introduced as a prototype for modelling complex disorders as quantitative traits.

Variance components and heritability: Here we demonstrate how phenotypic variance can be decomposed into genetic and non-genetic components, allowing the estimation of heritability (the fraction of phenotypic variance due to genetic variance). Methods for estimating heritability are discussed, including worked examples with inbred plant lines and monozygotic twins.

Dominance, additivity and interaction: Our genetic model for complex traits is further decomposed to explicitly define variance conferred by dominance effects, gene-gene and gene-environment interactions and additive effects of trait-increasing alleles. Methods for heritability estimation using non-twin pedigree data are defined.

Selection on complex traits and the liability scale for complex disorders: This lecture discusses the practical application of statistical models for complex traits, explaining models of variance components established in previous lectures through the lens of plant and animal breeding. The equivalence between this and the liability scale for complex disease is delineated.

Mapping complex disease genes I: GWAS and polygenic risk: Having established the overall model for complex diseases, this lecture now lays out a key method used to understand the genetics of complex disease: the genome-wide association study (GWAS). We also describe and explain how GWAS data can be used to understand the genetic architecture of complex diseases (e.g., polygenic/non-polygenic) and to estimate the fraction of overall heritability explained by common variants.

Mapping complex disease genes II: missing heritability and rare variants: This lecture defines the missing heritability problem along with explanations and solutions for understanding the genetic architecture of disease in the post-GWAS era, specifically in the context of rare variants.

We shift our focus from GWAS and rare single-nucleotide variants to some other types of genetic variation that are likely to play a major role in the aetiology of complex diseases. The lecture takes a dive into a worked example of the role of repeat expansions in the neurodegenerative disease amyotrophic lateral sclerosis.

8. Learning Outcomes:

On completion of the module students will:

- Have gained an overview of the history of medical genetics and understand the current state-of-the art in this rapidly changing and exciting field
- Have knowledge regarding the genetic basis of mendelian and complex disease
- Have gained insights into methods of analyses used in medical genetics; linkage studies, genome-wide association studies (GWAS), next generation sequencing (NGS), data analyses tools etc
- Understand the genetics of polygenic traits, the concept of heritability and methods to estimate it
- Have insights into emerging themes in personalized medicine that influence drug design, drug choice and drug response and the importance of genetic information for medicine and drug development
- Understand the relevance of medical genetics to diagnosis, prognosis and treatment of disease

9. Recommended Reading List:

'Genetic and Genomics and Medicine', (Garland Science) - Tom Strachan, Judith Goodship and Patrick Chinnery

'Introduction to Genetic Analysis' (Macmillan Education) Griffiths, Wessler, Carroll & Doebly

Primary references will be provided in lecture materials

10. Assessment Details: One 1.5-hour exam paper at the end of semester 2

11. Module Coordinator Jane Farrar gjfarrar@tcd.ie

BIU33250 Introduction to Immunology & Immunometabolism

Last year schedule and module descriptor. New timetable and details will be provided on Blackboard. Please use this information as a guide

- 1. Module Code** BIU33250 ([Open Module](#))
- 2. Module Name** Introduction to Immunology & Immunometabolism
- 3. Semester taught** Semester 2
- 4. Contact Hours** 22
- 5. Module Personnel** Cliona O'Farrelly (COF), Frederick Sheedy (FS), Jean Fletcher (JF), Richard Porter (RP), Luke O'Neill (LON),
- 6. Learning Aims** This module introduces to the basic components and function of the immune system – the molecules, cells, tissues and organs that make up the immune system. It will illustrate the immune responses to infection. Additionally, it will introduce students to the importance of central energy and intermediary metabolic pathways before considering how they are dysregulated in diseases like cancer and also how we can harness this knowledge for new immunotherapies
- 7. Module content: Provisional for semester 2**

Lecture #	Lecture Topic
23	Introduction – The Immune System (COF)
23	Innate Immunity 1 – Innate Defences (FS)
23	Innate Immunity 2 – Cellular Response to infection (FS)
23	Innate Immunity 3 – PRR Signalling (FS)
24	Innate Immunity 4 – Cytokines (FS)
24	T-cells 1 – DCs & Antigen Presentation (JF)
24	T-cells 2 – T-cell Receptor (JF)
24	T-cells 3 – T-cell Signalling (JF)
25	T-cells 4 – Effector T-cells (JF)
25	B-cells 1 – B-lymphocytes & Plasma Cells (COF)
25	B-cells 2 – Antibodies (COF)
25	B-cells 3 – Disorders of the immune system (COF)
26	Advanced Metabolism 1 – Central Energy Metabolism (RKP)
26	Advanced Metabolism 2 – Intermediary Metabolism (RKP)
26	Advanced Metabolism 2 – Intermediary Metabolism (RKP)
26	Advanced Metabolism 4 – Nucleotide Metabolism (RKP)
27	Advanced Metabolism 5 – Cancer Cell Metabolism (RKP)
27	Advanced Metabolism 6 – Immune Cell Metabolism (RKP)
27	Immunometabolism 1 (LON)
27	Immunometabolism 2 (LON)
28	In-class MCQ

Lecture schedule will be confirmed before start of semester 2.

8. Learning Outcomes:

On successful completion of this module students will be able to:

- Identify cells, receptors and soluble component of the innate immune system and how they function to eliminate pathogen.
- Define how an adaptive immune response is initiated and how different types of adaptive immune responses are used to eliminate particular pathogens.
- Identify how the immune system can cause disease and how it can be exploited therapeutically

- Recall key central energy and intermediary metabolic pathways and appreciate their importance in cellular function
- Apply knowledge on cellular metabolism to diseases including cancer and inflammation

9. Recommended Reading List:

The recommended text for this module is Janeway's Immunobiology published by Norton's Books, currently in its 10th Edition. Further reading will be given out by lecturers during the module.

10. Delivery

This Open module consists of 16 scheduled lectures (plus 2 tutorials) which will be delivered either online through Panopto (pre-recorded) or face to face. Any recordings will be released online in Blackboard and students will be notified beforehand. The in-class MCQ will be made available on the Blackboard after Reading/revision Week 1 at a flexible slot. The end of Semester exam will be an in-person, written exam paper.

11. Assessment Details:

60% End of year examination, 40% in course assessed.

In course assessment: In-class end of module MCQ exam covering lecture material

12. Module Coordinator **Dr Aisling Dunne** aidunne@tcd.ie

Last year schedule and module descriptor. New timetable and details will be provided on Blackboard. Please use this information as a guide

1. Module Code BIU33475 (Open Module) Module)

2. Module Name Basics of Neurobiology

3. Semester taught Semester 2

4. Contact Hours 16 hours (16 Lectures)

5. Module Personnel Gavin Davey & David Loane

6. Learning Aims: This module focuses on chemical transmission between neurons, how neurotransmitters are classified and identified and describes typical and atypical neurotransmitters and their functions in the brain. It considers mechanisms in which abnormal neurotransmission gives rise to common neurological & psychiatric disorders.

7. Module content: Programme of lectures and practicals –

Week	Lecture Topic & Lecturer
Semester	
22	Intro: cell types in the brain and their functions; NT types; NT criteria (GD)
22	Techniques for studying neurotransmission (GD)
22	Acetylcholine release & exocytosis (GD)
23	Biogenic Amines I (GD)
23	Biogenic Amines II & brain disorders (GD)
23	Glutamatergic neurotransmission systems
24	GABAergic neurotransmission systems (GD)
24	Atypical Neurotransmitters I (GD)
24	Atypical Neurotransmitters II (GD)
25	Brain lipids, gangliosides & lipid mediators (GD)
25	Intracellular trafficking & signalling (GD)
25	Inborn metabolic diseases of the brain (GD)
26	Inborn metabolic diseases of the brain (DL)
26	Neurobiology of schizophrenia & autism (DL)
27	Neurobiology of mood and anxiety disorders (DL)
27	Neurobiology of addiction (DL)

8. Learning Outcomes:

On completion of this module, the student will be able to:

- Describe the cell types in the brain and common techniques that enable chemicals with neurotransmitter-like properties to be identified
- Understand the criteria that need to be satisfied in order for a chemical to be classified as a neurotransmitter
- Develop a knowledge of the biogenic amines (acetylcholine, dopamine, noradrenaline, adrenaline, serotonin) and the properties that allow them to be classified as neurotransmitters
- Develop a knowledge of glutamate and GABA and the properties that allow them to be classified as neurotransmitters
- Develop a knowledge of atypical neurotransmitters (NO, CO, D-serine, neuropeptides, purines) and the properties that allow them to be classified as neurotransmitters
- Develop a knowledge of how dysfunctional neurotransmitter systems give rise to common neurological & psychiatric disorders

9. Recommended Reading List:

Basic Neurochemistry (Siegal, Albers, Brady, Price) Academic Press, 7th Edition. (6th Edition is online free at <https://www.ncbi.nlm.nih.gov/books/NBK20385/?term=basic%20neurochemistry>)

Principles of Neural Science by Eric Kandel , James Schwartz , Thomas Jessell , Steven Siegelbaum , A.J. Hudspeth

10. Assessment Details: Examination (70% written examinations; 30% continual assessment).

11. Module Coordinator: Dr Gavin Davey gdavey@tcd.ie

4.4 Marking Scale

Note that these guidelines are for use as a general reference. Differences may occur between disciplines.

	Mark Range	Criteria
I	90-100	IDEAL ANSWER; showing insight and originality and wide knowledge. Logical, accurate and concise presentation. Evidence of reading and thought beyond course content. Contains particularly apt examples. Links materials from lectures, practicals and seminars where appropriate.
	80-89	OUTSTANDING ANSWER; falls short of the 'ideal' answer either on aspects of presentation or on evidence of reading and thought beyond the course. Examples, layout and details are all sound.
	70-79	MAINLY OUTSTANDING ANSWER; falls short on presentation and reading or thought beyond the course, but retains insight and originality typical of first class work.
II-1	65-69	VERY COMPREHENSIVE ANSWER; good understanding of concepts supported by broad knowledge of subject. Notable for synthesis of information rather than originality. Sometimes with evidence of outside reading. Mostly accurate and logical with appropriate examples. Occasionally a lapse in detail.
	60-64	LESS COMPREHENSIVE ANSWER; mostly confined to good recall of coursework. Some synthesis of information or ideas. Accurate and logical within a limited scope. Some lapses in detail tolerated.
II-2	55-59	SOUND BUT INCOMPLETE ANSWER; based on coursework alone but suffers from a significant omission, error or misunderstanding. Usually lacks synthesis of information or ideas. Mainly logical and accurate within its limited scope and with lapses in detail.
	50-54	INCOMPLETE ANSWER; suffers from significant omissions, errors and misunderstandings, but still with understanding of main concepts and showing sound knowledge. Several lapses in detail.
III	45-49	WEAK ANSWER; limited understanding and knowledge of subject. Serious omissions, errors and misunderstandings, so that answer is no more than adequate.
	40-44	VERY WEAK ANSWER; a poor answer, lacking substance but giving some relevant information. Information given may not be in context or well explained, but will contain passages and words, which indicate a marginally adequate understanding.
F-1	35-39	MARGINAL FAIL; inadequate answer, with no substance or understanding, but with a vague knowledge relevant to the question.
	30-34	CLEAR FAILURE; some attempt made to write something relevant to the question. Errors serious but not absurd. Could also be a sound answer to the misinterpretation of a question.
F-2	0-29	UTTER FAILURE; with little hint of knowledge. Errors serious and absurd. Could also be a trivial response to the misinterpretation of a question.
U.G		Ungraded

Faculty of Engineering, Mathematics and Science - Guidelines on Marking, last modified 2007.

4.5 Attendance Requirements

Students must attend all practical and tutorial modules GEU33085, GEU33007, GEU33008, GEU33301 and are strongly advised to attend all lectures on the remaining modules. Failure to attend one third of the scheduled contact hours may deem the student non-satisfactory.

Our aim during the Junior Sophister year is to provide you with a thorough grounding in the fundamentals of modern Genetics so that you will be well prepared for the challenges of the Senior Sophister year. We therefore expect you to attend all lectures and in addition, some modules have strict attendance policy with direct consequences as a student may be deemed non-satisfactory.

Attendance at seminars

In addition to the lecture courses, there are weekly departmental seminars scheduled for 1 p.m. on Fridays in the atrium (these will be announced a few days ahead of time). Attendance is expected at these seminars.

Communication

Announcements will be made by emailing you at your **tcd.ie email address**. You must read this mailbox regularly or set it up to forward to an account that you do read. Your primary contact for each module is the module coordinator and for general queries, the Course Coordinator.

4.6 Non-submission of coursework and absence from Examinations

Students are required to complete the assessment components for each module as prescribed by the programme regulations.

Students must complete and submit the assessment components specified for each module that constitute their programme of study. Completion includes the submission of continuous assessment and attendance at examinations and other tests.

Students who are experiencing difficulties that are affecting their ability to complete their assessment components should contact their Tutor at the earliest opportunity to discuss the nature of the difficulties and the possible options available in Trinity. Depending on the specific details of a case, options can range from a request for a short extension from a module coordinator to a formal request for a deferral made to the Senior Lecturer/Dean of Undergraduate Studies.

Where the difficulties are serious, a student may need to make a Student Case, through their Tutor, to the Senior Lecturer/Dean of Undergraduate Studies.

Where a student does not complete specified assessment component(s), the relevant Court of Examiners has the authority to make one of the following determinations:

- i. Permission to defer to the reassessment session
- ii. Re-assess in relation to the missed component(s) for the reassessment session, subject to capping the associated reassessment(s) at the pass mark
- iii. Repeat year

Calendar Part II, B: General Regulations and Information, 'Absence' Academic Policies

4.7 Feedback and script viewing

Exam scripts can be viewed shortly after publication of results. Please contact genetics@tcd.ie to arrange script viewing.

4.8 Progression Regulation

It is important that you aim to achieve high grades in your continuous assessments and exams because 30% of the marks obtained in the JS year will contribute directly to your Senior Sophister BA Moderatorship grade. Also, when **project and review topics for Senior Sophister year** are assigned next year, students with higher marks in JS year will tend to get their higher preference choices of topic.

The current regulations are included below (calendar 2025-26). Please check the most recent version of the calendar for any updates ([TCD Calendar](#)):

Progression regulations: Bachelor programmes

59 Some programmes with professional accreditation have received a derogation from specific regulations on progression by the University Council. The relevant programme entry provides these details. See www.tcd.ie/academic-affairs/academic-regulations/-undergraduate-progression-and-awards.

In order to rise with their class, students must obtain credit for the academic year by satisfactory attendance at lectures and tutorials and by carrying out, submitting and sitting the required assessment components. In addition, students must pass the year by achieving, at a minimum, an overall credit-weighted average pass mark for the year (40 per cent or 50 per cent, as per programme regulations) and either:

- (a) accumulate 60 credits by achieving at least the pass mark in all modules or*
- (b) pass by compensation. All modules and components within modules are compensatable (except in particular professional programmes where compensation does not apply).*

To pass a year by compensation, in programmes that locate the pass mark at 40 per cent, a student must achieve the pass mark in modules carrying a minimum of 50 credits and obtain a module mark of at least 35 per cent in any remaining module(s). A student may accumulate a maximum of 10 credits at

qualified pass where the mark lies between 35-39 per cent.

To pass a year by compensation, in programmes that locate the pass mark at 50 per cent, a student must achieve the pass mark in modules carrying a minimum of 50 credits and obtain a module mark of at least 45 per cent in any remaining module(s). A student may accumulate a maximum of 10 credits at qualified pass where the mark lies between 45-49 per cent.

60 Progression is on an annual basis. Within a year students may carry failed modules from one semester to the next but not from one academic year to another; that is, they will not be able to rise to the next year of their programme until they have successfully completed the preceding year(s). Students who have not passed their year are required to present for reassessment when:

- (a) they obtain in excess of 10 credits at qualified pass (i.e. marks between 35-39 per cent where the pass mark is 40 per cent; or 45-49 per cent where the pass mark is 50 per cent);*
- (b) they fail any module (i.e. achieving marks below 35 per cent where the pass mark is 40 per cent; or below 45 per cent where the pass mark is 50 per cent);*
- (c) they do not obtain an overall pass mark for the year;*
- (d) any combination of (a) - (c) occurs.*

61 *If a student has achieved both fail and qualified pass grades at the first sitting or has exceeded the 10 credit limit allowed for compensation and is not*

permitted to rise with their year, they must present for reassessment in all modules for which they obtained a fail and/or a qualified pass.

62 *Different modalities of assessment to the first sitting are permitted in the reassessment session, as determined by the programme.*

63 *The same progression and compensation regulations as outlined above apply at the reassessment session. The overall credit-weighted average for the academic year will be calculated using the most recent marks achieved.*

64 *Students who fail to satisfy the requirements of their year at the reassessment session are required to repeat the year in full (i.e. all modules and all assessment components).*

65 *Students are permitted to repeat any year of an undergraduate programme subject to not repeating the same year more than once and not repeating more than two academic years within a degree course, except by special permission of the Senior Lecturer/Dean of Undergraduate Studies or the University Council.*

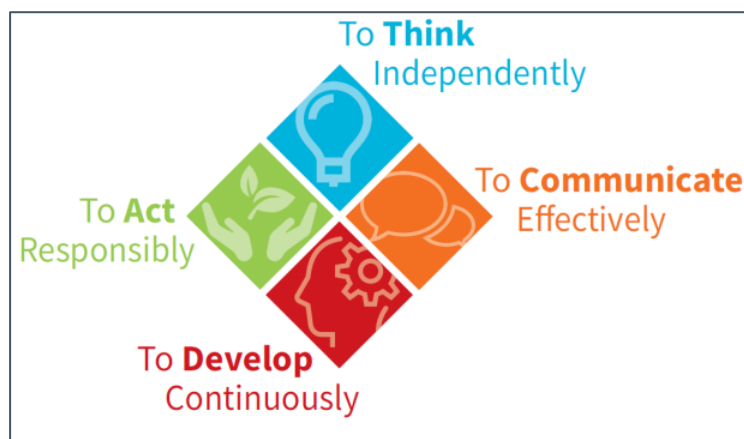
66 *The maximum number of years to complete an undergraduate degree is six years for a standard four-year programme and seven years for a five-year programme.*

There is one reassessment session which is held at the beginning of Michaelmas term. Students are assessed in all failed modules from both semesters during the reassessment session. Students are not permitted to repeat successfully completed assessments or examinations in order to improve their performance. In exceptional circumstances such as illness, if a student does not attempt exams at the end semester, they can defer until the reassessment examining period. Applications to defer exams should be made to the Senior Lecturer's Office via your tutor.

You are strongly advised to submit all the lab reports during the year, and not to miss the continuous assessment tests, because if you obtain a mark of < 35% in 3 or more modules you will be unable to progress to the Senior Sophister year (you'll have to repeat the JS year or leave College). If you fail to submit coursework on time, you will get a mark of zero for it.

4.10 Graduate Attributes

The Trinity Graduate Attributes represent the qualities, skills and behaviours that you will have the opportunity to develop as a Trinity student over your entire university experience, in other words, not only in the classroom, but also through engagement in co- and extra-curricular activities (such as summer work placements, internships, or volunteering).



The four Trinity Graduate Attributes are:

- To Think Independently
- To Act Responsibly
- To Develop Continuously
- To Communicate Effectively

Why are the Graduate Attributes important?

The Trinity Graduate Attributes will enhance your personal, professional and intellectual development. They will also help to prepare you for lifelong learning and for the challenges of living and working in an increasingly complex and changing world.

The Graduate Attributes will enhance your employability. Whilst your degree remains fundamental, also being able to demonstrate these Graduate Attributes will help you to differentiate yourself as they encapsulate the kinds of transversal skills and abilities, which employers are looking for.

How will I develop these Graduate Attributes?

Many of the Graduate Attributes are 'slow learned', in other words, you will develop them over the four or five years of your programme of study.

They are embedded in the curriculum and in assessments, for example, through undertaking independent research for your final year project, giving presentations and engaging in group work. You will also develop them through the co-curricular and extra-curricular activities. If you help to run a club or society you will be improving your leadership skills, or if you play a sport you are building your communication and team-work skills.

4.11 Student Feedback Evaluation

Annual undergraduate module evaluation - administered at School/course office level. Schools have discretion to choose which type of evaluation best suits their needs and to design their own questions. Schools are required to report on the type of evaluation conducted for each module, how it communicated feedback to students and actions arising from the evaluation.

5. Literature Review Instructions

Please note: due to the length and number of reviews to be assessed, we may not be able to return marks to you before the annual exams in May.

Format

Your review must not exceed 4,000 words. What is included in this limit? All text except: Title page, index page/table of contents, references section, appendixes and academic integrity page, page numbers, page header or footer.

What is included? All the rest including text in figure legends or within tables

Word limit is strictly enforced. However, a margin of 10% above the limit is allowed and there is no penalty below. Shorter, concise sentences are encouraged. Limit may not apply for some LENS students.

It must be typed in Times New Roman 12 point font, with a line spacing of 1.5. It must be submitted not later than Monday January 5th, 2026), with the word count verified and included in the submitted version (see appendix n IV).

- The work should be divided into: Title page; Abstract; Introduction; Main text to be organized in subsections with headings, according to topic; Conclusion and discussion; References. It may also contain an index/table of contents (encouraged)
- Pages must be numbered.
- Figures and Tables: If figures and tables are included, they must be numbered (Figure 1, etc.).
- Each Figure must be accompanied by an explanatory legend (text attached to the Figure that explains what it shows). If you have included a figure or table that you did not draw yourself, you must write your own legend and cite the source in the legend, e.g. "figure from (Jones et al 2018)". If you used a published figure and modified it cite the as "adapted from ...
- Each Figure/Table must be referred to from a sentence in the main text, to direct the reader to them when relevant.
- Citations: When the text refers to a published paper, the citations in the text must use a format like these examples (using EndNote APA 7th format should take care of everything):
 - XYZ was observed (Behan, 2011). For papers with 1 author.
 - XYZ was observed (Behan and Murphy, 2011). For papers with 2 authors.
 - XYZ was observed (Behan et al., 2011). For papers with 3 or more authors.
 - Most references are cited at the ends of sentences like this (Behan et al., 2011). However, it is sometimes more useful to write something like Behan et al. (2011) found that XYZ was not true.
 - Do not use a citation system based on numbers.
 - Do not include the initials of the authors in the citations in the main text.
- References section: The references section (also called the bibliography) is the list of papers that have been cited in the text. It appears at the end of the review. It gives more details of the papers that have been cited: complete list of authors (initials and surname); year of publication; title of the article; journal name; volume number; page numbers (first and last). Please use APA 7th format with EndNote as it will take care of all the formatting automatically. PLEASE DO NOT INCLUDE NUMBERS Example:
 - Behan M, Cahill S, Murphy C (2011) The plastid genome of higher plants. Nature Reviews Genetics 103: 56-58.

- References to websites should not be used as a substitute for the primary published literature in the field under review and should only be cited if there is no published paper as an alternative. If you need to cite a website, put the address (URL) directly in the text of as a footnote, not in the references section.
- The review's title and your name should be on the front cover.
- Make sure you are aware of College policies regarding academic misconduct (Appendix I, II and III) and complete the 'Ready, Steady, Write' online tutorial.
- Review submission is online and you must upload an electronic copy of your review to *Blackboard* before the deadline.

Other Review requirements:

WARNING: Your review must not contain material produced via a process of “copying” or “cutting and pasting” of text from any source: this is plagiarism. Also, the “paraphrasing” of text (i.e. changing few words in a sentence or paragraph) constitutes plagiarism. It can be readily detected by computer-based searches of your submitted work. For more information on plagiarism please see the appendix.

Plagiarism or use of AI where not authorized is a serious offence. Any submitted work (e.g. your Review) that contains content not adhering to College's academic integrity policy will be marked punitively and may even be awarded a mark of 0%.

Note that your supervisor will be able to gauge your level of understanding of the subject matter in your meetings with them, and this may be taken into account in their assessment of the final review. In addition, students may be invited to present their review and questioned on its content

Declaration information

Please include the declaration statement ([Appendix n IV](#)) on your review. Statement should be signed (electronic signature will suffice) and dated.

The review must be your own work. Note that your supervisor will be able to gauge your level of understanding of the subject matter in your meetings with them, and this may be taken into account in their assessment of the final review.

Review Assessment

The review will be assessed taking into account the following criteria:

- Difficulty of topic
- Scientific content
- Clarity of thinking / Comprehension of the subject
- Ability to write scientifically
- Structure of the review
- Awareness of the recent literature on this subject
- Presentation

6. Assessments guidelines

The overall Junior Sophister results will represent 30% of your final moderatorship grade. Modules are assessed by continuous assessment and/or by examination. The distribution scheme of marks between papers continuous assessment and practical work varies with each module and it is specified within its

description. Specific exam dates as well as submission deadlines are also specified for each module and it is vital that you submit on time.

- **Word limit:** Word limit is strictly enforced. However, a margin of 10% above the limit is allowed and there is no penalty below. **Shorter, concise answers are encouraged.** Limit may not apply for some LENS students.
- **Time:** The allocated time includes submission of the answers. A 5-10min lateness will be allowed without penalty. Some LENS students are allowed extra time and, although module coordinator will be aware, the student should make sure required accommodations are taken into account.
- **Submission:** Blackboard is the preferred submission platform. Submissions are allowed in the system past the deadline (they are flagged in red as late in Blackboard).

Penalties are applied when time or length are exceeded, and it only applies to the time or length beyond the allowed margin.

- **Exceeding the word limit:** Marker will consider when grading the ability of the student to adhere to the specified limit.
- **Exceeding the time limit:** Open book exams, 5% grade reduction/15min.
Assignments, review 5% grade reduction/day.

No penalties will apply if there are mitigating circumstances or the student has been allowed to exceed the limits. The onus is on the student to provide evidence of mitigating conditions. These incidences should be approved on a case-by-case basis by the Course Coordinator for consistency.

Mitigating circumstances are well documented technical problems. LENS students' particular circumstances or sickness properly justified.

7. The Library of Trinity College Dublin

History

Trinity College Library dates back to the establishment of the College in 1592. It is the largest library in Ireland with over 6 million printed volumes comprising extensive collections of books, journals, manuscripts, maps and music reflecting over 400 years of academic development. The Library was endowed with [Legal Deposit](#) privilege in 1801 and continues to receive material published in the United Kingdom and Ireland. Find out more at [About the Library](#) on The Library homepage.

Resources for Genetics Students:

The Library is currently composed of 7 different libraries. The ones most impactful to Genetics students are the Hamilton Library and the Library Complex, which contains the Lecky, Ussher, and Boland libraries. Books, articles, and databases can be accessed from the Library's website at www.tcd.ie/library/.

[My Library Top Tips](#)

1. Search the [Library Catalogue](#),
2. Contact their [Subject Librarian](#) for expert searching help. The subject librarian for Genetics is Leslie Kotzas (kotzasl@tcd.ie),
3. Borrow up to 10 books at a time,
4. Request books from the shelves or from storage using Click and Collect service in the library Catalogue,
5. Use the Self-Service Kiosks to borrow or return books,
6. Print/Copy/Scan on any library or campus printer (service managed by Datapac). You will need an ID code (your student number) and a PIN which is emailed to you when you register. You can change this PIN
7. Attend any training session - Library HITS - and brush up on your research skills,
8. Ask Library Staff for help by e-mail to library@tcd.ie, our chat with us on the Library website or in-person at our Service Desks.

More information on the Library can be found here: <https://www.tcd.ie/library/usinglibrary/>

Appendices

I. Online central repository

All students are required to access the **online central repository** in which all information and resources on plagiarism have been consolidated. This facility explains what Academic Integrity is, and how it can be avoided. The central repository is being hosted by the Library and is located at [About this guide - Academic Integrity - Library Guides at Trinity College Dublin](#)

About this guide

Introduction

What does good academic practice look like?

Study practices

Referencing practices

What is academic misconduct and how do I avoid it?

What are the consequences of academic misconduct?

Where can I go for further support?

Coversheet declaration

Ready Steady Write Plagiarism Tutorial

GenAI LibGuide

All students must read and understand this guide, and complete the [Ready Steady Write tutorial](#). We appreciate that it is sometimes challenging for students to recognise what good academic practice looks like. These resources are designed to help you understand and avoid academic misconduct as you develop your academic writing skills. Writing with academic integrity, that is, not plagiarising ideas or representing work you have not done yourself as your own, is a key expectation of Trinity students.

Looking for help with acknowledging or referencing the use of Generative AI? See our [Generative AI LibGuide](#).



Trinity's statement on Integrity

What does this mean to you as a student?

Why is academic integrity important?

What does academic integrity look like at Trinity?

II. Ready Steady Write Plagiarism Tutorial

All students are required to complete the **Ready Steady Write plagiarism tutorial**, a resource developed by the Centre for Academic Practice and eLearning (CAPSL) at Trinity College Dublin, to help you understand and avoid plagiarism and develop your academic writing skills and academic integrity.

www.tcd.ie/Library/support/plagiarism/story.html

Plagiarism can occur in many forms, for example copying another student's work, or quoting directly from published sources without acknowledgement, or using as your own slightly modified versions of the published work of others. Thus, in writing essays or other project work you are warned against copying verbatim, or copying and making minor modifications to, phrases, sentences, paragraphs, sections or illustrations from other published work.

Students and staff have access to Turnitin computer software (see Appendix III) that can readily detect plagiarism. The Department will use this sensitive anti-plagiarism tool to screen essays and other forms of formal assessed work and Turnitin reports can be used as evidence if plagiarism is suspected.

Accordingly, you are strongly recommended to synthesize your own language at all times. A full statement of the College's position on can be found in the College Calendar

III. Turnitin – Blackboard

Turnitin is an online software program that aids plagiarism prevention. It allows students and lecturers to check students' work for academic integrity by searching for text that is improperly cited or potentially plagiarised. Once uploaded to Turnitin, assignments are compared to millions of books, journal articles, web pages and student papers, identifying any unoriginal material within the essay. The software then creates an Originality Report which highlights and quantifies unoriginal content.

For more information, see <http://tcd-ie.libguides.com/plagiarism/detecting-plagiarism> and to access the student training tutorial, see http://www.turnitin.com/en_us/training/student-training

IV. Declaration to be included on literature review

TRINITY COLLEGE DUBLIN THE UNIVERSITY OF DUBLIN

SCHOOL OF GENETICS AND MICROBIOLOGY

SMURFIT INSTITUTE OF GENETICS

DECLARATION FOR REVIEW

I have read and I understand the academic integrity provisions in the General Regulations of the University Calendar for the current year, found at <http://www.tcd.ie/calendar>.

I have also completed the Online Tutorial 'Ready Steady Write', located at <https://libguides.tcd.ie/academic-integrity/ready-steady-write>

I declare that this assignment fully complies with
College's academic integrity provisions.

Signed.....

Dated

The **word count** of this document (with the exception of
the References section is:

Signed.....

Dated

V. Academic year structure 2026/27

Academic calendar week	Week beginning	2026/27 Academic Year Calendar		Term / Semester
		UG continuing years / PG all years	UG new first years	
1	31-Aug-26	Marking/Results 2025/26		←Michaelmas Term begins/Semester 1 begins
2	07-Sep-26	Marking/Results and Orientation (PG, Visiting, Erasmus)		
3	14-Sep-26	Teaching and Learning	Orientation (IF UG)	←Michaelmas teaching term begins
4	21-Sep-26	Teaching and Learning	Teaching and Learning	
5	28-Sep-26	Teaching and Learning	Teaching and Learning	
6	05-Oct-26	Teaching and Learning	Teaching and Learning	
7	12-Oct-26	Teaching and Learning	Teaching and Learning	
8	19-Oct-26	Teaching and Learning	Teaching and Learning	
9	26-Oct-26	Study/Review (Monday, Public Holiday)	Study/Review (Monday, Public Holiday)	
10	02-Nov-26	Teaching and Learning	Teaching and Learning	
11	09-Nov-26	Teaching and Learning	Teaching and Learning	
12	16-Nov-26	Teaching and Learning	Teaching and Learning	
13	23-Nov-26	Teaching and Learning	Teaching and Learning	
14	30-Nov-26	Teaching and Learning	Teaching and Learning	
15	07-Dec-26	Revision / Assessment*	Revision / Assessment*	
16	14-Dec-26	Assessment*	Assessment*	←Michaelmas Term ends Sunday 20 December 2026/Semester 1 ends
17	21-Dec-26	Assessment* / Christmas	Assessment* / Christmas	
18	28-Dec-26	Christmas Period - College closed 24 December 2026 to 3 January 2027 inclusive	Christmas Period - College closed 24 December 2026 to 3 January 2027 inclusive	
19	04-Jan-27	Foundation Scholarship Examinations	Foundation Scholarship Examinations	
20	11-Jan-27	Marking***	Marking***	
21	18-Jan-27	Teaching and Learning	Teaching and Learning	←Hilary Term begins/Semester 2 begins; Hilary teaching term begins
22	25-Jan-27	Teaching and Learning	Teaching and Learning	
23	01-Feb-27	Teaching and Learning (Monday, Public Holiday)	Teaching and Learning (Monday, Public Holiday)	
24	08-Feb-27	Teaching and Learning	Teaching and Learning	
25	15-Feb-27	Teaching and Learning	Teaching and Learning	
26	22-Feb-27	Teaching and Learning	Teaching and Learning	
27	01-Mar-27	Study/Review	Study/Review	
28	08-Mar-27	Teaching and Learning	Teaching and Learning	
29	15-Mar-27	Teaching and Learning (Wednesday, Public Holiday)	Teaching and Learning (Wednesday, Public Holiday)	
30	22-Mar-27	Teaching and Learning (Friday, Good Friday)	Teaching and Learning (Friday, Good Friday)	
31	29-Mar-27	Teaching and Learning (Monday, Easter Monday)	Teaching and Learning (Monday, Easter Monday)	
32	05-Apr-27	Teaching and Learning	Teaching and Learning	
33	12-Apr-27	Revision	Revision	
34	19-Apr-27	Assessment**	Assessment**	←Hilary Term ends Sunday 25 April 2027
35	26-Apr-27	Trinity Week (Monday, Trinity Monday) / Assessment**	Trinity Week (Monday, Trinity Monday) / Assessment**	←Trinity Term begins
36	03-May-27	Marking/Results (Monday, Public Holiday)	Marking/Results (Monday, Public Holiday)	
37	10-May-27	Marking/Results	Marking/Results	
38	17-May-27	Marking/Results	Marking/Results	
39	24-May-27	Research	Research	
40	31-May-27	Research	Research	←Trinity Term ends Sunday 6 June 2027/Semester 2 ends
41	07-Jun-27	Research (Monday, Public Holiday)	Research (Monday, Public Holiday)	
42	14-Jun-27	Research	Research	
43	21-Jun-27	Research	Research	
44	28-Jun-27	Research	Research	
45	05-Jul-27	Research	Research	
46	12-Jul-27	Research	Research	
47	19-Jul-27	Research	Research	
48	26-Jul-27	Research	Research	
49	02-Aug-27	Research (Monday, Public Holiday)	Research (Monday, Public Holiday)	
50	09-Aug-27	Research	Research	
51	16-Aug-27	Research	Research	
52	23-Aug-27	Reassessment 2026/27 - Semesters 1 & 2	Reassessment 2026/27 - Semesters 1 & 2	

* Semester 1 assessment session: December 11 to 22, 2026 inclusive (No assessment after Dec 22nd)

** Semester 2 assessment session: April 19 to 30, 2027 inclusive

*** Marking of Semester 1 assessments will continue into January and early February. Provisional Semester 1 results will be made available to students during the week commencing February 8, 2027

VI. Study Abroad declaration

Personal Declaration for Study Abroad School of Genetics and Microbiology

Please read this declaration carefully, in full, before signing and submitting your application

Study abroad does not suit everyone. Before applying, students should reflect carefully on their individual academic goals, learning styles, personal circumstances etc.

1. You should have confidence in answering 'YES' to the following statements:

- I am aware of the academic and personal challenges that can arise during study abroad.
- I am not aware of any specific issue(s) or factor(s) that would adversely affect my ability to adapt to a different learning environment or to integrate smoothly into a different cultural environment.
- (where applicable) I realise that I will have to adapt to a different linguistic environment and am confident in my language abilities.

2. Respect for host University:

Exchange students must follow the rules of their host university, particularly with regard to subject assessment and examinations, and accept that conditions may differ from those in Trinity.

I understand that:

- it is my responsibility to inform the host University of any needs I may have.
- the School of Genetics and Microbiology in Trinity College Dublin cannot guarantee accommodations abroad.
- the School of Genetics and Microbiology in Trinity College Dublin cannot review the decisions of partner universities.

3. Compliance with institutional, national, EU and international regulations:

- College-Wide exchanges: I understand that my exchange is governed by the International Admissions and Study Abroad Office, while I am subject to School of Genetics and Microbiology rules, I am aware that my point of contact is the International Admissions and Study Abroad Office and that the School of Genetics and Microbiology merely facilitates this process.
- European exchanges: I understand and accept that study abroad is governed by regulations and procedures set out in ERASMUS and other relevant literature published by the EU, the Higher Education Authority (Ireland) and other relevant national and supranational bodies.

4. Conversion of Marks

I understand that the School of Genetics and Microbiology converts grades in the fairest way possible, with great concern for its student's grades and future prospects. I accept that conditions in my host university may differ from those in Trinity and realise that this may have an effect on my results.

Please note that the School of Genetics and Microbiology does its utmost to ensure that the translation of results from your host university to Trinity's marking system is fair and reflects your achievements abroad. The Court of Examiners cannot, however, guarantee that the process of translation of marks will equal out all differences between the marking structures in different countries and amongst the host universities. The School of Genetics and Microbiology International Committee bases its conversions on conversion tables but retains a discretion to ensure broad equivalence. Trinity's conversion tables are attached to this document.

Individual marks are also carefully reviewed prior to sending them to the Court of Examiners. The distribution of grades in the host and home university may also be taken into account (i.e. your relative position in the class may also be considered), as well as the course load taken.

5. Meeting degree programme requirements

Prior to leaving for exchange, I have conducted extensive research on the institutions I have selected.

- I am aware of my course requirements in terms of credits and have discussed the same with the exchange coordinator(s) for all of my subjects.

Within two weeks of arriving at my host University I will:

- Email a scanned copy of my Learning Agreement, signed by my TCD coordinator(s) and host University to my programme administrator.

While on exchange I will:

- Obtain the permission of my coordinator(s) for any changes I am proposing to my learning agreement while away and resubmit my signed learning agreement to my programme administrator
- Inform my coordinators immediately on any difficulties, including failing modules, that may impact on my ability to progress

Prior to submitting this application, I have conducted extensive research on the institutions I have selected.

- I am aware of all of the information and course requirements which are highlighted in the Erasmus and Exchange Rules 2026/27.
- I understand that I am going abroad as a Trinity student. I therefore understand that I am an ambassador for the College and for the School of Genetics and Microbiology. I will bear this in mind in all my conduct abroad, both academic and personal.
- I am aware I must obtain not less than 25 ECTS credits, or equivalent, from the host university while studying abroad during semester two.
- I confirm that I have read and accept in full the content of this declaration and I confirm that the information provided by me in the application form for study abroad is correct.

(Please type your name below if you are submitting the document electronically)

Name: _____ Date: _____

VII. Table conversion

European Erasmus Conversion Table for Trinity Students Returning from Study Abroad

IRELAND Trinity College	F 0 – 39	III 40 – 49	II.2 50 – 59	II.1 60 – 69	I 70 +
ECTS	Fail	E-D	D-C	B	A
AUSTRIA	5 Nicht genügend	4 Genügend	3 Befriedigend	2 Gut	1 Sehr Gut
BELGIUM	0 – 9	10 – 11	12 – 13	14 – 15	16+
CZECH REPUBLIC*	4	3**	3**	2	1
DENMARK	-3 – 0	02	4	7	10 – 12
FINLAND (2 scales)	0	1 – 2 1 – 1.5	2 – 3 2	3 – 4 2.5	4 – 5 3
FRANCE	0 – 7	8 – 9	10 – 12	13 – 15	16+
GERMANY	6.0 – 4.7	Noch ausreichend – Noch Befriedigend 4.5 – 3.7	Befriedigend to Noch Gut 3.5 – 2.7	Gut to Sehr Gut 2.5 – 1.7	Sehr Gut to Ausge zei chnet 1.5 – 1.0
GREECE	0 – 5	5 – 5.9	6 – 6.9	7 – 8.4	8.5+
ICELAND (ECCITS & Flexi Scale)	0-4.99 0-4.99	5.00-5.99 5.00-6.00	6.00-7.24 6.00-7.5	7.25-8.99 7.5-8.5	9.00-10.00 8.5-10.00
ITALY	0 – 17	18 – 22	23 – 26	27 – 29	30 – 30 e lode
MALTA	F 0 – 44	D – D+ 45 – 54	C – C+ 55 – 69	B – B+ 70 – 79	A – A + 8 0 +
NETHERLANDS	0 – 5.4	5.5 – 6.4	6.5 – 7.0	7.1 – 7.9	8.0+
POLAND	0 – 2	3 – 3.5	4	4.5	5 – 5!
PORTUGAL	0 – 9	10 – 11	12 – 13	14 – 15	16+
SPAIN	Suspense 0 – 4.9	Aproba do 5 – 5.9	6 – 6.9	Notable 7 – 8.9	Sobresal i ente 9 Sobresal iente Sobresal iente 10 Matricula de honor (80+)

SWEDEN	U 0 – 49	G** 50 – 55	G** 56 – 70	VG** 71 – 90	V G * * 9 0 +
SWITZERLAND	0 – 2.75	3 – 3.5	3.75 – 4.25	4.5 – 5.5	5.75 – 6
TURKEY	0 – 49	50 – 64	65 – 74	75 – 84	85+
UK (England, Northern Ireland & Wales)	F 0 – 39	D- – D+ 40 – 49	C- – C+ 50 – 59	B- – B+ 60 – 69	A - -
					A + 7 0 +
UK (Scotland) UK (St Andrews, Scotland)	0 – 8 0 – 6	9 – 11 7 – 10	12 – 14 11 – 13	15 – 17 14 – 16	18 – 20 17 – 20
RUSSIA	2	3	4	5 (Excellent)**	5 (Excellent)**
Important: 1. The aim of the conversion table is to ensure that all results are treated equally by all disciplines. 2. If for exceptional reasons a discipline deviates from the table, they must inform the International Admissions and Study Abroad Office in writing, stating the reasons for the deviation. Approved deviations include the School of Business (for Business Schools in France only) and the School of Law which uses more refined university by university grades conversion tables in line with the above grade categories. 3. Official transcripts issued by the host institution supersede any unofficial grades supplied by students.					
Footnotes: * Charles University in Prague might provide more detailed results on request. Students should take a copy of the Trinity grading scheme with them and present it to individual lecturers on arrival as agreed with the International Office in Prague. ** Where a mark spans two Trinity grades the higher grade may be awarded where supporting documentation is received.					

VIII. Prizes and other awards

RONALD A. FISHER PRIZE IN GENETICS The Ronald A. Fisher prize in genetics is awarded annually to a Sophister student who has excelled in oral presentation of a topic of his/her own choice within the field of genetics. The prize was established by a gift from George Dawson who founded the Department of Genetics in 1958 and led it until 1987. Sir Ronald Fisher, Baldwin Professor of Genetics at Cambridge (1943-57), was acknowledged as a leading authority on genetical theories of natural selection and statistics and was a major influence on George Dawson, a student at Cambridge. The prize will be awarded on the advice of the Head of the Department of Genetics.

Value, **€191**

DAWSON PRIZE IN GENETICS This prize was founded in 1990 by colleagues and friends of George Dawson, member of staff (1950-87) and Professor of Genetics (1967-87). He founded the department in 1959 and started a programme of summer research for rising Senior Sophisters. The prize will be awarded, on the advice of the Professor of Genetics, to the student of genetics who has distinguished himself/herself in the Junior Sophister examinations.

Value, **€316**

A.W.B. VINCENT SCHOLARSHIPS IN GENETICS AND HUMAN GENETICS These scholarships are awarded to students in genetics and human genetics, selected by competition, to facilitate research internships at institutions for two to three months in the summer of their Junior Sophister year. William Vincent, LL.D. (h.c.), (1919-2012) established these scholarships in 1975 and funded them for 40 years. His family and friends contributed to ensure they would continue after his death. Dr Vincent was president of the American Irish Foundation from 1972 to 1987 when it merged with the American Ireland Fund of which he became vicechairman. He was a generous supporter of many charitable projects in Ireland.

Value, **€1500**

Larry O'Hara

The Larry O'Hara scholarship will facilitate JS Genetics or Human Genetics students to work in a laboratory in Ireland or internationally for the summer in between JS and SS years. The student should make contact with a laboratory that would be willing to host them and then send details of the laboratory and the proposed project to genetics@tcd.ie. A committee chaired by the Head of Discipline will then review all applications and award the scholarship(s) accordingly. Applications must be submitted by the end of February.

IX. Safety statement

To ensure the health and safety of everyone in the Genetics department, we will share with you the [departmental Safety Statement](#). **We ask you to read and abide by the rules given in the Safety Statement.** Please note that failure to comply with the procedures outlined in the departmental Safety Statement may result in disciplinary action. The information provided below is for convenience only; it does not substitute for the departmental Safety Statement.

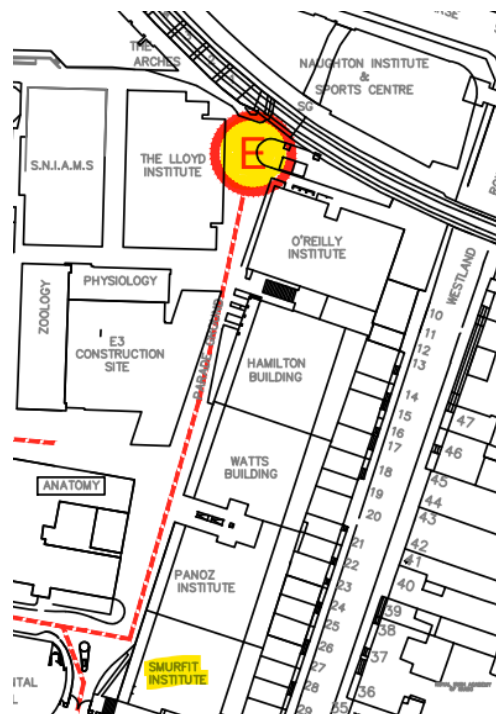
Here we highlight the most relevant safety rules concerning undergraduate students who enter laboratories and lecture theatres. These rules must be followed at all times.

General

- Students are not allowed to enter laboratories unless they are authorized to do so.
- Students are not permitted to work in laboratories unsupervised.
- Students must always follow the instructions of laboratory supervisors.
- Eating and drinking is prohibited in laboratories and lecture theatres.
- Smoking is strictly prohibited in all campus buildings.
- Do not leave coats, bags or personal belongings on lab benches or anywhere where they could cause obstruction.
- Students should not congregate at the entrance to a lab or lecture theatre or at building entrances.

Fire safety

- In the event of a fire alarm,
LEAVE THE BUILDING immediately using the nearest exit route.
- Report to the Genetics Department Assembly point 'E' shown on the map here, between the O'Reilly / Lloyd buildings.



Laboratory dress code

- Wear long trousers or skirts, and shoes with non-slip soles that fully cover your feet. Shorts, open-toed sandals, flip-flops, high heels, ballet pumps, crocs, and canvas shoes/runners are not permitted.
- Students must wear a suitable laboratory coat while working in a laboratory. “Howie-style” laboratory coats are preferred.
- Safety glasses must be worn when there is access/use of chemicals or potential exposure to biological agents containing aerosols. Those wearing spectacles must wear ‘Pulsafe’ kind which are worn over the normal spectacles. Contact lenses may constitute an additional hazard.
- Long hair must be properly tied back and adequately restrained.
- No loose hanging jewellery, headphones, or earbuds permitted in the laboratory.
- Gloves must be worn and changed as required in all laboratory environments involving the use of chemicals or biological agents.

Laboratory safety

- If any glass apparatus/container/pipette breaks while in use, inform a member of staff immediately.
- Ensure caps are replaced on all containers with chemicals when an experiment is completed.
- If you come into direct contact with chemicals, inform a member of staff immediately.
- Familiarize yourself with the location of first aid kits, safety showers, and eye wash stations in the laboratory you are working in.
- Students must familiarize themselves with the European Standard Chemical hazard symbols shown below.

 Gas under pressure Symbol: gas cylinder	 Explosive Symbol: exploding bomb	 Oxidising Symbol: flame over circle
 Flammable Symbol: flame	 Corrosive Symbol: corrosion	 Acute Health Hazard Symbol: flame over circle
 Acute toxicity Symbol: skull and crossbones	 Serious Health Hazard Symbol: health hazard	 Hazardous to the environment Symbol: environment

If you have any concerns about Health and Safety in the department, please contact the departmental Safety Officer (Sr. Technical Officer - Orla Deevy) or the Head of Department (currently Prof. Matthew Campbell). For further information on Health and Safety, see the website of the College Safety office at <https://www.tcd.ie/safetyoffice/>

X. Careers Advisory Service

What do you want to do? How will you get there? We are here to support you in answering these and other questions about your career.

Junior and Senior Fresh Students

Get Involved: Remember that your course of study, extra-curricular activities, voluntary and part-time work all provide opportunities for developing skills and gaining an insight into your career preferences. In your Senior Fresh year, look out for short-term internship opportunities.

MyCareer: Log in to MyCareer to keep abreast of jobs, study and careers events of interest to you.

Junior Sophisters

Attend class seminar: Typically this takes place in Hilary term and includes information on applying for work experience and internships and postgraduate study.

Get work experience: The programme of summer work experience and internships is particularly relevant to Junior Sophisters. Personalise your MyCareer profile to receive email alerts tailored to your preferences.

MyCareer: Log in to MyCareer to keep abreast of jobs, study and careers events of interest to you.

Finalists and Senior Sophisters

Meet Employers and/or Explore Further Study: You may have decided to seek employment directly after graduation and many employers visit Dublin to actively seek out talented graduates. For others, further study may be their preferred option. Your

MyCareer dashboard will keep you informed.

Find Jobs: Personalise your MyCareer profile to receive email alerts tailored to your interests.

Attend class seminar: Typically this takes place in Michaelmas term and includes information on applying for postgraduate study and jobs.

GradLink Mentoring: An opportunity to get advice and support from a Trinity graduate.

Drop-In CV/ LinkedIn Clinics: We also provide support at a practical level, helping you to improve your applications, which will benefit you in securing your future, whether in employment or further study.

Practice Interviews: A practice interview tailored to the job/ course of your choice with practical feedback.

MyCareer: Log in to MyCareer to keep abreast of jobs, study and careers events of interest to you.

MyCareer

An online service that you can use to:

- Apply for opportunities which match your preferences - vacancies including research options
- Search opportunities- postgraduate courses and funding
- View and book onto employer and CAS events
- Submit your career queries to the CAS team
- Book an appointment with your Careers Consultant

Simply login to MyCareer using your Trinity username and password and personalise your profile.

Careers Advisory Service

Trinity College Dublin, 7-9 South Leinster Street, Dublin 2
01 896 1705/1721 | Submit a career query through [MyCareer](#)

 mycareerconnect.tcd.ie

 [TCD.Careers.Service](#)

 [TCDCareers](#)

 www.tcd.ie/Careers/students/postgraduate

 [@TCDCareers](#)

 [tinyurl.com/LinkedIn-TCD-Connecting](#)

Opening Hours

During term: 9.30am - 5.00pm, Monday - Friday
Out of Term: 9.30am - 12.30pm & 2.15 - 5.00pm, Monday - Friday